

CHEMICAL CONSTITUENTS OF *CAESALPINIA BONDOC*

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Four known triterpenoids (1-4) α -amyrin (1), β -amyrin (2), lup-20(29)-en-3 β -ol (3), lup-20(20)-en-3 β -yl acetate (4) and two known sterols (5,6) β -sitosterol (5) and β -sitosteryl galactoside (6) were isolated first time from the *Caesalpinia bonduc*.

Key words: *Caesalpinia bonduc*, Triterpenoids,

Introduction

The seeds of *Caesalpinia bonduc* (L.) Roxb. belonging to the family Caesalpinaceae were used as a folk medicine to treat asthma and chronic fever. They were found to be very potent as an antiperiodic, antipyretic and utilized as febrifuge, antispasmodic, antirheumatic and mild purgative [1-2]. Literature survey revealed the presence of diterpenoids such as α -caesalpin [3-4], β -caesalpin [3,4], δ -caesalpin [4,5], γ -caesalpin [3-6] and ϵ -caesalpin [7] from the seeds of *Caesalpinia bonduc* (L.) Roxb. [syn. *C. bonducella* (L.) Fleming]. Our phytochemical investigations have led to the isolation of four triterpenoids α -amyrin [12-ursen-3 β -ol] (1), β -amyrin [12-oleanen-3 β -ol] (2), lup-20(29)-en-3 β -ol (3), lup-20(29)-en-3 β -yl acetate (4), and two sterols β -sitosterol [24(R)-stigmast-5-en-3 β -ol] (5), β -sitosteryl galactoside [3-O- β -D-galactopyranosyl-stigmasta-5-ene] (6).

Experimental

The seeds of *Caesalpinia bonduc* were collected from Karachi during the months of November-December. The fresh seeds (6.5 kg) were ground in a homogenizer and soaked in MeOH for two weeks. It was filtered and the filtrate was concentrated under reduced pressure. The concentrated crude extract (95 g) was partitioned between EtOAc and H₂O. The aqueous layer was again shaken with *n*-BuOH. The EtOAc extract was condensed by removing the solvent under reduced pressure and the material thus obtained was subjected to column chromatography using solvent systems *n*-hexane; *n*-hexane; CHCl₃; CHCl₃; CHCl₃: MeOH.

Compound 1 was eluted with *n*-hexane: CHCl₃ (9:1) and crystallized from EtOH, yielded 40 mg of 1.

α -Amyrin (1): Mp 184-185°C; [α]_D + 90° (CHCl₃; C, 0.30); EIMS *m/z* (rel. intens. %): 426 [C₃₀H₅₀O, M⁺] (20), 411 [M-Me]⁺ (16), 408 [M-H₂O]⁺ (25), 393 [M-Me-H₂O]⁺ (22), 257 [M-C₁₁H₂₁O]⁺ (20), 218 [M-C₁₄H₂₄O]⁺ (100), 207 [M-C₁₆H₂₇O]⁺ (12), 203 [M-C₁₅H₂₇O]⁺ (50) and 189 [M-C₁₆H₂₉O]⁺

(65); IR (CHCl₃) ν_{\max} cm⁻¹: 3510 (OH), 3055, 1635 and 820 (trisubstituted double bond); ¹H-NMR (300 MHz, CDCl₃) [8]: δ 5.23 (1H, m, H-12), 3.21 (1H, dd, J_{ax,ax}} = 10.2 Hz, J_{ax,eq}} = 4.3 Hz, H-3), 1.15 (3H, s, Me-27), 1.01 (3H, s, Me-26), 0.99 (3H, s, Me-23), 0.95 (3H, s, Me-25), 0.88 (6H, s, Me-29 and Me-30), 0.83 (3H, s, Me-28) and 0.80 (3H, s, Me-24); ¹³C-NMR (75 MHz, CDCl₃) [9]: 838.51 (C-1), 27.00 (C-2), 77.00 (C-3), 37.99 (C-4), 55.10 (C-5), 18.32 (C-6), 32.65 (C-7), 40.00 (C-8), 47.56 (C-9), 37.00 (C-10), 23.00 (C-11), 122.52 (C-12), 143.50 (C-13), 41.52 (C-14), 28.32 (C-15), 26.23 (C-16), 32.52 (C-17), 47.20 (C-18), 46.81 (C-19), 31.12 (C-20), 34.81 (C-21), 37.20 (C-22), 26.01 (C-23), 15.56 (C-24), 15.50 (C-25), 16.80 (C-26), 26.00 (C-27), 27.32 (C-28), 33.20 (C-29) and 23.69 (C-30).

The eluent obtained with *n*-hexane: CHCl₃ (9:1, v/v) was again subjected to silica gel column chromatography to obtain 2 which was crystallized from EtOH, yielded 30 mg of 2.

β -Amyrin (2): Mp 197-198°C; [α]_D + 99° (CHCl₃; C, 0.40); EIMS *m/z* (rel. intens. %): 426 [C₃₀H₅₀O, M⁺] (15), 411 [M-Me]⁺ (18), 408 [M-H₂O]⁺ (16), 393 [M-Me-H₂O]⁺ (32), 257 [M-C₁₁H₂₁O]⁺ (20), 218 [M-C₁₄H₂₄O]⁺ (100), 207 [M-C₁₆H₂₇O]⁺ (10), 203 [M-C₁₅H₂₇O]⁺ (40) and 189 [M-C₁₆H₂₉O]⁺ (55); IR (CHCl₃) ν_{\max} cm⁻¹: 3510 (OH), 3055, 1635 and 820 (trisubstituted double bond); ¹H-NMR (300 MHz, CDCl₃) [9]: δ 5.11 (1H, m, H-12), 3.19 (1H, dd, J_{ax,ax}} = 10.0 Hz, J_{ax,eq}} = 4.5 Hz, H-3), 1.08 (3H, s, Me-27), 1.02 (3H, s, Me-26), 1.01 (3H, s, Me-23), 0.96 (3H, s, Me-25), 0.91 (3H, d, J=6.6 Hz, Me-30), 0.81 (6H, s, Me-24) and 0.80 (3H, d, J=6.8 Hz, Me-29); ¹³C-NMR (75 MHz, CDCl₃) [10]: 839.00 (C-1), 27.31 (C-2), 78.99 (C-3), 39.00 (C-4), 55.23 (C-5), 18.30 (C-6), 33.00 (C-7), 41.01 (C-8), 47.80 (C-9), 37.00 (C-10), 17.44 (C-11), 124.30 (C-12), 139.34 (C-13), 42.00 (C-14), 28.71 (C-15), 26.62 (C-16), 33.76 (C-17), 59.00 (C-18), 39.61 (C-19), 39.56 (C-20), 31.20 (C-21), 41.52 (C-22), 28.12 (C-23), 15.62 (C-24), 15.95 (C-25), 16.81 (C-26), 23.32 (C-27), 28.00 (C-28), 23.39 (C-29) and 21.34 (C-30).

The fraction eluted with *n*-hexane: CHCl₃ (9:1, v/v) was finally purified by re-column chromatography using *n*-hexane

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: CHCl_3 (8.5 : 1.5, v/v) and crystallized from Me_2CO -MeOH, yielded 100 mg of 3.

Lupeol (3): Mp 214-215°C; $[\alpha]_D + 27^\circ$ (CHCl_3 ; C, 1.00); EIMS m/z (rel. intens. %): 426 [$\text{C}_{30}\text{H}_{50}\text{O}$, M^+] (20), 411 [M-Me] $^+$ (25), 408 [M-H $_2\text{O}$] $^+$ (30), 393 [M-Me-H $_2\text{O}$] $^+$ (35), 385 [M-41] $^+$ (15), 220 [M-C $_{15}\text{H}_{26}$] $^+$ (80), 218 [M-C $_{14}\text{H}_{24}\text{O}$] $^+$ (55), 207 [M-C $_{16}\text{H}_{27}$] $^+$ (25) 189 [M-C $_{16}\text{H}_{29}\text{O}$] $^+$ (100) and 139 [M-C $_{21}\text{H}_{35}$] $^+$ (70); IR (CHCl_3) ν_{max} cm^{-1} : 3455 (OH), 3075, 1645 and 880 (exomethylene group); $^1\text{H-NMR}$ (300 MHz, CDCl_3) [10]: δ 4.75 and 4.62 (2H, br. s, 1H each, H-29), 3.21 (1H, dd, $J_{\text{ax,ax}} = 9.9$ Hz, $J_{\text{ax,cq}} = 4.5$ Hz, H-3), 1.65 (3H, br. s, Me-30), 1.05 (3H, s, Me-26), 0.97 (3H, s, Me-23), 0.94 (3H, s, Me-27), 0.85 (3H, s, Me-25) 0.79 (3H, s, Me-28) and 0.77 (3H, s, Me-24); $^{13}\text{C-NMR}$ (75 MHz, CDCl_3) [10]: δ 838.62 (C-1), 27.55 (C-2), 78.81 (C-3), 38.76 (C-4), 55.33 (C-5), 18.30 (C-6), 34.25 (C-7), 40.85 (C-8), 50.44 (C-9), 37.11 (C-10), 20.95 (C-11), 25.20 (C-12), 38.10 (C-13), 42.80 (C-14), 27.39 (C-15), 35.51 (C-16), 92.91 (C-17), 48.22 (C-18), 47.79 (C-19), 150.65 (C-20), 29.89 (C-21), 39.88 (C-22), 28.02 (C-23), 15.42 (C-24), 16.11 (C-25), 15.99 (C-26), 14.52 (C-27), 18.11 (C-28), 109.25 (C-29) and 19.20 (C-30).

The fraction obtained by *n*-hexane : CHCl_3 (1 : 1, v/v) was subjected to repeated chromatographic purifications using *n*-

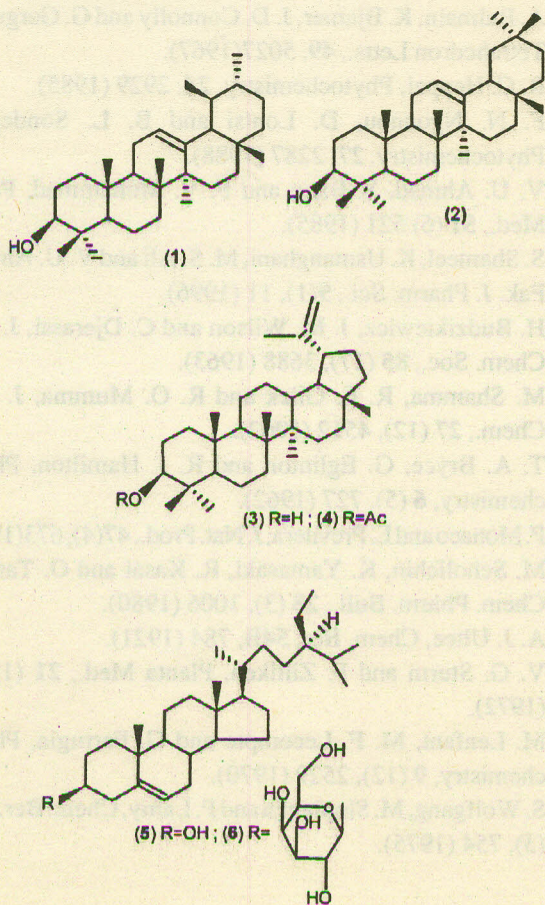
hexane : EtOAc (9 : 1, v/v) afforded compound 4 which was crystallized from Me_2CO : MeOH (yield: 32 mg).

Lupeol Acetate (4): Mp 214-216°C; $[\alpha]_D + 41^\circ$ (CHCl_3 ; C, 0.32); EIMS m/z (rel. intens. %): 468 [$\text{C}_{23}\text{H}_{32}\text{O}_2$, M^+] (50), 453 [M-Me] $^+$ (15), 427 [M-C $_3\text{H}_5$] $^+$ (5), 408 [M-AcOH] $^+$ (20), 393 [(453)-AcOH] $^+$ (3), 249 [M-C $_{16}\text{H}_{27}$] $^+$ (22), 218 [M-C $_{16}\text{H}_{26}\text{O}_2$] $^+$ (37), 189 [(249)-AcOH] $^+$ (63), 181 [M-C $_{21}\text{H}_{35}\text{O}$] $^+$ (10) and 121 [(181)-AcOH] $^+$ (48); IR (CHCl_3) ν_{max} cm^{-1} : 1710 (ester carbonyl) and 3075, 1645 and 880 (exomethylene group); $^1\text{H-NMR}$ (400 MHz, CDCl_3) : δ 4.72 and 4.62 (2H, br. s, 1H each, H-29), 4.25 (1H, dd, $J_{\text{ax,ax}} = 9.8$ Hz, $J_{\text{ax,cq}} = 4.2$ Hz, H-3), 2.11 (3H, s, CH_3COO), 1.65 (3H, dd, $J = 1.25$ Hz, Me-30), 1.05 (3H, s, Me-26), 0.97 (3H, s, Me-23), 0.94 (3H, s, Me-27) 0.85, (3H, s, Me-25) 0.79 (3H, s, Me-28) and 0.77 (3H, s, Me-24); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) : δ 38.41 (C-1), 23.75 (C-2), 81.05 (C-3), 37.81 (C-4), 55.43 (C-5), 18.22 (C-6), 34.30 (C-7), 40.91 (C-8), 50.43 (C-9), 37.11 (C-10), 21.06 (C-11), 25.12 (C-12), 38.10 (C-13), 42.94 (C-14), 27.52 (C-15), 35.65 (C-16), 43.08 (C-17), 48.07 (C-18), 48.31 (C-19), 150.95 (C-20), 29.99 (C-21), 40.00 (C-22), 28.05 (C-23), 16.51 (C-24), 16.22 (C-25), 16.03 (C-26), 14.50 (C-27), 18.06 (C-28), 19.33 (C-29), 109.41 (C-30), 21.32 (CH_3COO) and 170.81 (CH_3COO).

Compound 5 which was eluted with *n*-hexane : CHCl_3 (8 : 2, v/v) was finally purified by crystallization from MeOH (yield : 50 mg).

β -Sitosterol (5): Mp 134.5°C; $[\alpha]_D + 40^\circ$ (CHCl_3 ; C, 0.50); EIMS m/z (rel. intens. %): 414 [$\text{C}_{29}\text{H}_{50}\text{O}$, M^+] (30), 399 [M-Me] $^+$ (10), 396 [M-H $_2\text{O}$] $^+$ (15), 381 [M-Me-H $_2\text{O}$] $^+$ (75), 329 [M-H $_2\text{O}$ -C $_7\text{H}_7$] $^+$ (26), 303 [M-H $_2\text{O}$ -C $_7\text{H}_9$] $^+$ (20), 275 [M-H $_2\text{O}$ -C $_9\text{H}_{13}$] $^+$ (14), 273 [M-side chain at C-17] $^+$ (35) and 255 [M-side chain-H $_2\text{O}$] $^+$ (35); IR (CHCl_3) ν_{max} cm^{-1} : 3450 (OH), 3050, 1650 and 815 (trisubstituted double bond); $^1\text{H-NMR}$ (300 MHz, CDCl_3) [11]: δ 5.23 (1H, m, H-6), 3.32 (1H, m, H-3), 1.01 (3H, s, Me-19), 0.92 (3H, d, $J = 6.2$ Hz, Me-21), 0.84 (3H, t, $J = 7.0$ Hz, Me-29), 0.83 (3H, d, $J = 6.5$ Hz, Me-26), 0.81 (3H, d, $J = 6.5$ Hz, Me-27) and 0.68 (3H, s, Me-18), $^{13}\text{C-NMR}$ (75 MHz, CDCl_3) [11]: δ 837.31 (C-1), 31.81 (C-2), 71.90 (C-3), 42.40 (C-4), 140.90 (C-5), 121.87 (C-6), 32.07 (C-7), 32.00 (C-8), 50.81 (C-9), 36.61 (C-10), 21.12 (C-11), 40.00 (C-12), 42.61 (C-13), 56.78 (C-14), 24.32 (C-15), 28.24 (C-16), 56.20 (C-17), 11.90 (C-18), 19.44 (C-19), 36.26 (C-20), 19.10 (C-21), 34.00 (C-22), 29.31 (C-23), 45.80 (C-24), 26.21 (C-25), 18.80 (C-26), 19.80 (C-27), 23.10 (C-28) and 11.92 (C-29).

The eluate obtained from CHCl_3 : MeOH (9 : 1, v/v) was further purified through flash chromatography using various ratios of CHCl_3 and MeOH. The aliquot obtained by pure CHCl_3 was finally purified by washing with Me_2CO and crystallized from mixture of CHCl_3 and MeOH to get 6 (yield : 45 mg).



***β*-Sitosterol Galactoside (6)**: Mp 275-277°C; $[\alpha]_D + 63^\circ$ (Pyridine; C, 0.52); EIMS *m/z* (rel. intens. %) : 577 [M^+ , (M+H), FAB-MS⁺], Aglycone : 414 [$C_{29}H_{50}O^+$] (20), 396 [$M-H_2O^+$] (15), 381 [$M-Me-H_2O^+$] (26), 371 [$M-C_3H_7^+$] (35), [$M-C_3H_7-H_2O^+$] (75), 329 [$M-H_2O-C_5H_7^+$] (55), 303 [$M-C_7H_9-H_2O^+$] (20), 302 [$M-C_8H_{16}^+$] (65), 275 [$M-H_2O-C_9H_{13}^+$] (35); IR $\nu_{max} cm^{-1}$: 3440-3410 (OH), 3025, 1650 (C=C) and 1440 (gemdimethyls); ¹H-NMR (300 MHz, CDCl₃) [11] : δ Aglycone : 5.23 (1H, m, H-6), 3.72 (1H, m, H-3), 1.01 (3H, s, Me-19), 0.92 (3H, d, J = 6.2 Hz, Me-21), 0.84 (3H, t, J = 7.0 Hz, Me-29), 0.83 (3H, d, J = 6.5 Hz, Me-26), 0.81 (3H, d, J = 6.5 Hz, Me-27) and 0.68 (3H, s, Me-18). Sugar : 4.20 (1H, d, J = 7.8 Hz, H-1), 3.65 (1H, dd, J = 1.0, 3.0 Hz, H-4) 3.55 (1H, dd, J = 8.0, 9.0 Hz, H-2), 3.52 (1H, dd, J = 7.0, 11.0 Hz, H-6), 3.30 (1H, dd, J = 5.0, 11.0 Hz, H-6), 3.15 (1H, dd, J = 3.0, 9.0 Hz, H-3) and 3.03 (1H, dd, J = 1.0, 5.0 Hz, H-5); ¹³C-NMR (75 MHz, CDCl₃) [11] : δ Aglycone : 37.31 (C-1), 31.81 (C-2), 78.85 (C-3), 42.40 (C-4), 140.90 (C-5), 121.87 (C-6), 32.07 (C-7), 32.00 (C-8), 50.81 (C-9), 36.61 (C-10), 21.12 (C-11), 40.00 (C-12), 42.61 (C-13), 56.78 (C-14), 24.32 (C-15), 28.24 (C-16), 56.20 (C-17), 11.90 (C-18), 19.44 (C-19), 36.26 (C-20), 19.10 (C-21), 34.00 (C-22), 29.31 (C-23), 45.80 (C-24), 26.21 (C-25), 18.80 (C-26), 19.80 (C-27), 23.10 (C-28) and 11.92 (C-29). Sugar : 100.51 (C-1), 71.90 (C-2), 74.00 (C-3), 69.90 (C-4), 76.53 (C-5) and 62.58 (C-6).

Acid hydrolysis of 6: Compound 6 (12 mg) was hydrolysed with 2N HCl in dilute MeOH (9mL methanol + 1 mL H₂O) at 100° for 3 hrs. The solvent of the reaction mixture was evaporated under reduced pressure. The residue was diluted with water and neutralized with Ag₂CO₃. The reaction mixture was then extracted with ethylacetate to furnish the aglycone moiety, while the aqueous layer was evaporated under reduced pressure. The residue obtained showed the presence of *β*-D-galactose when compared with the authentic sample by TLC using the solvent system EtOAc : MeOH : AcOH : H₂O (6 : 2 : 1 : 1). The spots were detected by spraying with sugar reagent (Solution A: 1gm FeCl₃ in 100 ml of 10% H₂SO₄ ; Solution B: 6% orcin in EtOH. Solutions 1 & 2 were mixed in the ratio of 10:1, respectively).

Results and Discussion

The four triterpenoids (1-4) α -amyrin [12-ursen-3 β -ol] (1) [12-14], β -amyrin [12-oleanex-3 β -ol] (2) [12-13], lup-20(29)-en-3 β -ol (3) [10, 12, 16-17], lup-20(29)-en-3 β -ylacetate (4) [17-19] and two sterols (5,6) β -sitosterol [24(R)-stigmast-5-en-3 β -ol] (5) [19-20] and β -sitosteryl galactoside [3-O- β -D-galactopyranosyl-stigmasta-5-enen] (6) [21] were isolated from the said source. Compounds 1-5 were isolated from the ethylacetate soluble part of the methanolic extract

whereas compound 6 was isolated from the butanolic part of the methanolic extract of the seeds of *Caesalpinia bonduc*. These compounds were first time isolated from the source but these have been already reported in the literature, so, they were not discussed extensively in the Results and Discussion section. They were just identified by direct comparison with published data [12-20].

References

1. G. V. Satiyavati, M. K. Raina and M. Sharma, *Medicinal Plants of India*, (Indian Council of Medicinal Res., New Delhi, 1976), Vol. I, pp.159.
2. K. M. Nadkarni and A. K. Nadkarni, *Indian Materia Medica*, (Bombay Popular Prakashan, 1976), Vol. I, pp.226.
3. M. Qudrat-i-Khuda and M.E. Ali, *Pak. j. sci. ind. res.*, **6**, 65 (1963).
4. A. Balmain, J. D. Connolly, M. Ferari, E. L. Ghisalberty, U. M. Pagoni and F. Pelizzoni, *Chem. Commun.*, **19**, 1244 (1970).
5. K. K. Purushothaman, K. Kalyani, K. Subramanian and S. Shanmuganathan, *Indian, Chem.*, **20B**, (7), 625 (1981).
6. C. Luigi, J. Giancarlo, M. Paolo, M. P. Ugo and P. Francesca, *Gazz. Chim. Ital.*, **96** (5), 662 (1966).
7. A. Balmain, K. Bjamer, J. D. Connolly and G. Gerguson, *Tetrahedron Letts.*, **49**, 5027(1967).
8. R. C. Heupel, *Phytochemistry*, **24**, 2929 (1985).
9. F. N. Ngounou, D. Lontsi and B. L. Sondegam, *Phytochemistry*, **27**, 2287 (1988).
10. V. U. Ahmad, S. Bano and F. V. Mohammad, *Planta Med.*, **51** (6) 521 (1985).
11. S. Shameel, K. Usmanghani, M. S. Ali and V. U. Ahmad, *Pak. J. Pharm. Sci.*, **9**(1), 11 (1996).
12. H. Budzikiewicz, J. M. Wilson and C. Djerassi, *J. Am. Chem. Soc.*, **85** (17), 3688 (1963).
13. M. Shamma, R. E. Glick and R. O. Mumma, *J. Org. Chem.*, **27** (12), 4512 (1962).
14. T. A. Bryce, G. Eglinton and R. J. Hamilton, *Phytochemistry*, **6** (5), 727 (1962).
15. P. Monaco and L. Previtera, *J. Nat. Prod.*, **47**(4), 673(1984).
16. M. Scholichin, K. Yamasaki, R. Kasai and O. Tanaka, *Chem. Pharm. Bull.*, **28** (3), 1006 (1980).
17. A. J. Ultee, *Chem. Ber*, **54B**, 784 (1921).
18. V. G. Sturm and F. Zilliken, *Planta Med.*, **21** (1), 61 (1972).
19. M. Lenfant, M. F. Lecompte and G. Farrugia, *Phytochemistry*, **9** (12), 2529 (1970).
20. S. Wolfgang, M. Slopianka and P. Lamy, *Chem. Ber.*, **108** (3), 754 (1975).