

# Technology Section

Pa. j. sci. ind. res., vol. 36, no. 4, April 1993

## A RAPID ASSAY PROCEDURE FOR THE DETERMINATION OF THIAMINE IN PHARMACEUTICAL PRODUCTS

M. AMINUDDIN AND M. SALEEM BHUTTA

*Chemistry Department, Islamia University, Bahawalpur, Pakistan*

(Received October 13, 1992; revised March 15, 1993)

A fast and simple method of determining thiamine (vitamin B<sub>1</sub>) in neat solution and in pharmaceutical products has been developed using titrimetric procedure. The method makes use of the reaction in which thiamine is oxidized [1] by potassium ferricyanide in thiochrome in an alkaline medium. In the present investigation the ferrocyanide ion produced in equivalent amount has been titrated in presence of the remaining excess potassium ferricyanide with zinc sulphate in an acid medium. The determination is interference free from the other vitamins, e.g., B<sub>2</sub>, B<sub>6</sub> and B<sub>12</sub>. Detection as low as 0.01 mg ml<sup>-1</sup> has been obtained. The time required for analysis is short, being about 5 mins for a single determination.

**Key words :** Thiamine, Assay, Pharmaceutical products.

### Introduction

It is well known that the determination of thiamine at small concentration levels is difficult, especially in food and pharmaceutical products. In order to remove the interference from the other B-vitamins, there has been applied numerous ways and means. So far it has not been possible to determine the vitamin B<sub>1</sub> without time-consuming separations [2-4], precipitation [5], or extraction based on either spectrophotometry or fluorimetry methods [6-10].

The interference from other soluble B-vitamins has been a major problem associated with all these methods. More recently developed techniques for the determination of vitamin B<sub>1</sub> in pharmaceutical preparations or a complex sample have included microbial [11], high performance liquid chromatography (HPLC) [12-14], spectrophotometry [15], spectrophotometry coupled with thin layer chromatography (TLC) [16]. Normally, however, these techniques have again demanded the separation of the samples as a preparatory treatment and it has involved considerable time and purchase of expensive apparatus. During our present study, we considered indirect titrimetric method, in which the vitamin is oxidized with potassium ferricyanide and the reduced form of oxidant in presence of potassium ferricyanide is titrated with zinc sulphate, to be an interesting alternative. Our main emphasis has been the applicability of the procedure to determine vitamin B<sub>1</sub> at very low concentration levels without involving separation process but being totally interference free from the other soluble B-vitamins. The sensitivity and simplicity of this method allow it to be useful for any routine analysis.

### Experimental

(a). *Materials and reagents.* All reagents were of analytical - reagent grade and doubled distilled water was used to

prepare all solutions. Vitamin B<sub>1</sub> (Fluka), Berin tablet and injection (Glaxo), Thianeuron capsule (Pfizer) and B-complex syrup Plexovit (Remington) were used in the present studies.

i. *Potassium ferricyanide solution (0.4 mM).* A suitable quantity of potassium ferricyanide was accurately weighed and dissolved in distilled water to obtain known volume of a 0.4mM solution.

ii. *Zinc sulphate (0.6 mM).* A known quantity of zinc sulphate (ZnSO<sub>4</sub> · 8H<sub>2</sub>O) was weighed accurately and dissolved in known volume of distilled water to obtain 0.6 mM solution.

iii. *Sodium hydroxide solution (0.1N).* 0.4 g of sodium hydroxide was dissolved in distilled water to make 100 ml solution.

iv. *Diphenylamine. (0.16%).* 0.16 g of Diphenylamine was dissolved in concentrated sulphuric acid and the volume was made upto 100 ml with the acid in a measuring flask.

(b). *Vitamin(s) solution.* i. *Vitamin B<sub>1</sub> neat solution.* 0.5g of vitamin B<sub>1</sub> was dissolved to make 500 ml solution in distilled water including a few millilitres of 0.1 N sodium hydroxide to maintain the solution at pH 8.5. By diluting this fifty times with distilled water a solution with 0.02 mg ml<sup>-1</sup> vitamin was obtained.

ii. *Mixture of vitamin B<sub>1</sub>, B<sub>2</sub> and B<sub>6</sub>.* Equal quantities of vitamin B<sub>1</sub>, B<sub>2</sub> and B<sub>6</sub> (0.5 g each) were dissolved in water to make 500 ml solution. 10 Millilitre of the solution was diluted to 500 ml using distilled water and a few millilitre of 0.1 N sodium hydroxide (to maintain pH 8.5). Each millilitre of the solution contains 0.02 mg each of B<sub>1</sub>, B<sub>2</sub> and B<sub>6</sub>.

### SAMPLE TREATMENT

i. *Berin tablet.* Two tablets equivalent to 200 mg of



TABLE 1. EFFECT OF pH ON THE DETERMINATION OF THIAMINE USING  $ZnSO_4$ , 0.0006M, AS TITRANT.

pH of the medium	Thiamine (mg)	$ZnSO_4$ 0.0006M (ml)	Corrected amount (Found, mg)	±Of Difference
7.5	0.02	0.1	0.0199	0.50
	0.04	0.2	0.0399	0.25
	0.08	0.3	0.0599	25.12
	0.12	0.4	0.0798	33.50
8.0	0.02	0.1	0.0199	0.50
	0.04	0.2	0.0399	0.25
	0.08	0.4	0.0798	0.25
	0.12	0.6	0.1196	0.33
8.5	0.02	0.1	0.0199	0.50
	0.04	0.2	0.0399	0.25
	0.08	0.4	0.0798	0.25
	0.12	0.6	0.1196	0.33
9.0	0.02	0.07	0.0139	30.50
	0.04	0.10	0.0199	50.25
	0.08	0.30	0.0598	25.25
	0.12	0.50	0.0997	16.91

*Role of acid and oxidant strength for successful titration.*

The reaction is thought to involve titration of ferrocyanic acid in presence of potassium ferricyanide with zinc sulphate. The medium thus needed to be acidified. A quantity 0.5 ml each of concentrated sulphuric acid and ortho-phosphoric acid was found to give sharper end point with 0.6 mM zinc sulphate using diphenylamine indicator. The use of concentrated sulphuric acid in larger quantity (1 ml or over) affected in the location of end point due to blackening of the titrating solution.

The strength of potassium ferricyanide was also found to affect the results. The procedure worked well with 0.4 mM potassium ferricyanide but as the strength was increased to 0.1M there was obtained a precipitate during titration with zinc sulphate. The procedure was thus restricted to the use of very dilute solution of the oxidant.

*Concentration effect.* In order to determine the range of concentration for which the procedure is applicable, experiments were carried out using pure vitamin  $B_1$  solutions of 0.010, 0.020 and 0.05  $mg\ ml^{-1}$  strengths. Table 2 illustrates interesting points regarding titration results of the vitamin at different concentrations. It was observed that scattering of the results was more associated with vitamins at higher concentration (0.05  $mg\ ml^{-1}$ ). The scattering of the results was completely absent for solutions studied at lower concentrations, e.g., 0.01-0.0  $mg\ ml^{-1}$ . This suggests that the method is suitable for smaller quantities determination. However, concentration of 0.02  $mg\ ml^{-1}$  of vitamin was selected for all further investigations.

TABLE 2. CONSUMPTION OF TITRANT ( $ZnSO_4$ , 0.0006M) AS A FUNCTION OF THE AMOUNT OF THIAMINE AT ITS DIFFERENT CONCENTRATIONS.

Thiamine $mg\ ml^{-1}$	Size of thiamine solution (ml)	$ZnSO_4$ 0.0006M (ml)	Thiamine (mg) Expected	Thiamine (mg) Found	±% Difference
0.010	1.0	0.05	0.010	0.0099	1.00
	2.0	0.10	0.020	0.0199	0.50
	3.0	0.15	0.030	0.0299	0.33
	4.0	0.20	0.040	0.0399	0.25
	5.0	0.25	0.050	0.0499	0.20
	6.0	0.30	0.060	0.0599	0.16
	7.0	0.35	0.070	0.0699	0.14
	8.0	0.40	0.080	0.0799	0.12
0.020	1.0	0.10	0.020	0.0199	0.50
	2.0	0.20	0.040	0.0399	0.25
	3.0	0.30	0.060	0.0599	0.20
	4.0	0.40	0.080	0.0799	0.12
	5.0	0.50	0.100	0.0987	1.30
	6.0	0.60	0.120	0.1198	0.16
	7.0	0.70	0.140	0.1398	0.14
	8.0	0.80	0.160	0.1595	0.31
0.050	1.0	0.20	0.050	0.0399	20.20
	2.0	0.45	0.100	0.0899	10.10
	3.0	0.65	0.150	0.1298	13.40
	4.0	0.80	0.200	0.1590	20.50
	5.0	1.00	0.250	0.2109	15.60
	6.0	1.25	0.300	0.2497	7.60
	7.0	1.40	0.350	0.2797	20.00
	8.0	1.55	0.400	0.3096	22.60

*Application.* The procedure has been evaluated by applying it to the determination of vitamin  $B_1$  in tablets, capsules, injectables, syrups and laboratory made mixtures. Detection in the lower concentration range 0.01 to 0.02  $mg\ ml^{-1}$  was a practical proposition. Table 3 presents results of the vitamin  $B_1$  in pharmaceutical products determined singly or in the presence of other B-vitamins, e.g.,  $B_2$ ,  $B_6$ , nicotinamide and  $B_{12}$ . The percentage difference in the determination is negligible, hardly being more than 0.01%. It is concluded that the procedure is reliable and free from interference not only by the B-vitamins but also the excipients and lubricants present in the preparations. Table 4 are the results of vitamin  $B_1$  obtained from the laboratory made mixture. The results further illustrated that the presence of other vitamins did not interfere in the determination of vitamin  $B_1$  and the procedure is quantitative, reproducible and interference free.

In conclusion the method is novel in the sense that the reaction is simple and free from any side reaction as far as the analysis of pharmaceutical products is concerned.

TABLE 3. QUANTITATIVE DETERMINATION OF VITAMIN B<sub>1</sub> IN PHARMACEUTICAL PRODUCTS.

Products	Label claim/ composition	Found quantity (mg)	Difference ±%
"Berin" Tablet (Glaxo)	Each tablet contains: Thiamine (100 mg)	99.99	0.01
"Berin" Injection (Glaxo)	Each ampoule contains: Thiamine (100 mg)	100.00	0.00
"Thianeuron" capsule (Pfizer)	Each capsule contains: Thiamine mononitrate (100 mg) Pyridoxine hydrochloride (200 mg), Cyanocobalamine (200 µg)	100.00	0.00
"Plexovit" Syrup (Remington)	Each 15 ml contains: Thiamine hydrochloride (3mg), Riboflavin (3 mg) Psyridoxine hydrochloride (2 mg), and Nicotinamide (23 mg)	2.99	0.0

TABLE 4. QUANTITY OF VITAMIN B<sub>1</sub> DETERMINED IN A LABORATORY MADE MIXTURE CONTAINING B<sub>1</sub>, B<sub>2</sub> AND B<sub>6</sub>.

Theoretical quantity (mg)	Found quantity (mg)	±% Difference
0.02	0.02	0.0
0.04	0.04	0.0
0.06	0.06	0.0
0.08	0.08	0.0
1.20	1.20	0.0

## References

1. I. Barger, *Ber.*, **68**, 2257 (1935).
2. N.P. Sen and A.D. Robinson, *J. Biochem. Physiol.*, **41**, 97 (1963).
3. M. Amin and J. Reusch, *Analyst*, **112**, 989 (1987).
4. G.R. Gerigh and H. Silberman, *Australian J. Pharm.*, **44**, 69 (1963).
5. D.C.M. Adamson and F.P. Hanisyde, *Quart. J. Pharm.*, **21**, 370 (1948).
6. G. Popova, *Tr. Nauch. Vissh. Inst. Khranit Vkusova Prom. Plodiv.*, **14**, 261 (1967).
7. H. Ogawara, Y. Horaguchi, M. Moroi and N. Nishimura, *Jpn. Kokai Tokyo Koho Jp*, 62 58, 166 (1987).
8. M. Berary, M. Abdul Hamedi, E. Hassan and M. Elsayed, *Pharmazie (Eng.)*, **41**, 483 (1986).
9. A.A. Moussa and S.M. Hassan, *Pharmazie*, **32**, 50 (1977).
10. D. Pearson, *The Chemical Analysis of Food*, (Churchill Livingstone, London, 1976), 7th ed., pp. 217.
11. M. Pallavicini and C. Redaelli, *Ann. Microbiol. Enzimol. (Italy)*, **37**, 7 (1987).
12. M. Metschies and G. Schwedt, *Dtsch. Lebensm Rundsch.*, (Ger), **81**, 385 (1985).
13. I.H. Pathan, J. Bartos, J. Zyka and A.M. Khawaja, *Sb.vys.sk.zemed. Praze, Pak. Agron, Rada A*, (Eng), **A-44**, 259 (1986).
14. N. Nakahara, Okada and H. Isaka, *Iyaku hin, Kenkyu*, (Japan), **19**, 84 (1988).
15. B.E. Burmudez, *Rev. Cubana Farm.*, **19**, 50 (1985).
16. G. Bengt, B. Halen and H. Johnson, *vaar Foeda (Swed)*, **41**, 51 (1989).