

GUAIANIN N, A NEW SAPONIN FROM FLOWERS OF *GUAIAECUM OFFICINALE*

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A new triterpenoidal saponin, guaianin N, (1) has been isolated from the butanolic extract of the flowers of *Guaiacum officinale*. It showed antibacterial activity against *Pseudomonas pseudomaliae* as well as brine shrimp toxicity [1]. Spectroscopic methods have been used to characterize compound 1 as 3-O-[β-D-glucopyranosyl (→3)α-L-arabinopyranosyl]-oleanolic acid.

Key words: Guaianin N, Saponin, *Guaiacum officinale*, Zygophyllaceae.

Introduction

The flower extract of *Guaiacum officinale* L. has been subjected to intensive investigation. The butanolic extract has shown significant activity against both gram positive and gram negative bacteria as well as toxicity against shrimps [1].

Guaiacum officinale L. (Zygophyllaceae) is both a medicinal as well as an ornamental plant and has been introduced into Pakistan from Africa and West Indies [2]. Both its resin and wood possess medicinal and pharmacological applications [3].

A number of saponins have already been reported from bark and leaves of *Guaiacum officinale*. The new oleanolic acid saponin guaianin N, 1, isolated from the flowers of *Guaiacum officinale* L. resembles Akebia saponin P_E [4] and Scabioside B isolated from *Patrinia scabiofolia* [5] with the difference in the linkages between the two sugars. The former has (1→2) and the latter has (1→4) linkage between the two sugars glucose and arabinose while 1 has (1→3) linkage between them.

Experimental

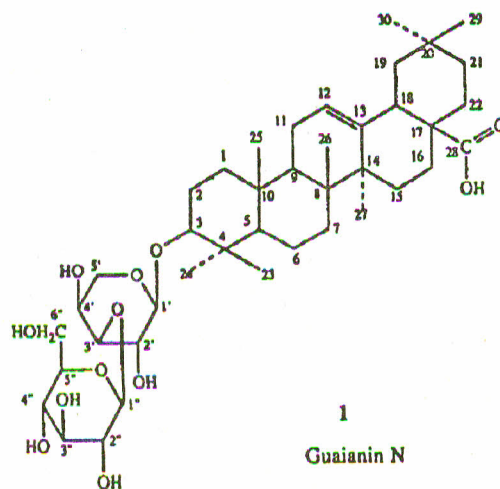
Melting point was recorded on Gallenkamp microscope. The ¹H-NMR and ¹³C-NMR spectra were recorded in CD₃OD on Bruker Aspect AM-3000 spectrometer at 300 MHz and 75.43 MHz respectively. Mass spectrum was recorded on JEOL, JMS, HX-110 connected with DA 5000 (Data system LS1 11/73 (CPU) DEC. E. Merck, Kiesel gel 60 GF₂₅₄ Art. 5719 glass plates were used for TLC analysis. Shimadzu liquid chromatography 6A HPLC was used for final purification.

Collection. The flowers of *Guaiacum officinale* L. were collected from the premises of Karachi University.

Extraction and isolation. 5 kilogram (wet wt.) of the flowers were crushed in methanol by Ultraturex homogenizer (Janke & Kunkel Co., Stanfen, Germany) and were extracted. The methanolic extract was evaporated under reduced pressure. The extractive was dissolved in water and then shaken with chloroform to remove terpenes and chlorophyll etc. The remaining water soluble extract was shaken with butanol and the butanolic extract was also evaporated under reduced

pressure and subjected to column chromatography on silica gel. The column was eluted with chloroform and with chloroform : methanol with increasing polarity. The fractions isolated with 7% methanol were subjected to repeated silica gel column chromatography and flash chromatography and finally guaianin N was isolated via HPLC using semi-preparative RP-18 column and differential refractometer detector. The solvent system used was 20% water in methanol; m.p.: 255-265° (decomposed); [α]_D: + 140 (c=0.27, MeOH); ¹H-NMR : δ0.84 (s, CH₃), 0.84 (s, CH₃), 0.90 (s, CH₃), 0.94 (s, CH₃), 0.94 (s, CH₃), 1.05(s, CH₃) and 1.09 (s, CH₃) indicating seven tertiary methyl groups. The anomeric signals at δ4.28 (d, J_{1,2}=7.1 Hz, H-1') and 4.50 (d, J_{1,2}=7.5 Hz, H-1'') show 1,2-diaxial relationship which is consistent for α-L-arabinopyranosyl and β-D-glucopyranosyl respectively and a distorted triplet at δ5.23 (t, H-12) indicates endocyclic double bond. ¹³C-NMR data of a glycone moiety is given in Table 1 while that of the two sugar moieties β-D-glucopyranosyl-(1→3)-α-L-arabinopyranosyl is given in Table 2 FAB-MS (negative mode high field) m/z 749 (M-H)⁻, C₄₁H₇₈O₁₂, 587 (M-glucose-H)⁻, 455 (M-glucose-arabinose-H)⁻.

Acid hydrolysis. 5 mg of compound 1 was refluxed with methanolic HCl (9ml MeOH, 1ml H₂O and 1.5 ml HCl) for 3 hrs. The reaction mixture was concentrated under reduced



pressure to remove methanol. It was then diluted with water, extracted with ethyl acetate. The aqueous layer was concentrated at reduced pressure and the residue obtained was compared with standard sugar on silica gel TLC. The solvent system for TLC was *n*-butanol : ethyl acetate : 2-propyl alcohol : acetic acid in 8.75 : 25 : 15 : 8.75 : 7.5 ratio. TLC card 20 cm long was run twice. Sugars spots were detected by spraying sugar reagent (orcinol + H₂SO₄ + FeCl₃). It also showed that sugars in compound 1 were arabinose and glucose.

Results and Discussion

The ¹H-NMR (C₅D₅N) exhibited signals at δ0.83 (s, 3 x H-25), 0.95 (s, 3 x H-29), 0.96 (s, 3 x H-24), 0.98 (s, 3 x H-26), 1.00 (s, 3 x H-30), 1.29 (s, 3 x H-23) and 1.30 (s, 3 x H-27) indicated 7 methyl group while the ¹H-NMR (CD₃OD) exhibited signals at δ0.84 (s, CH₃), 0.84 (s, CH₃), 0.90 (s, CH₃), 0.94 (s, H₃), 0.94 (s, CH₃), 1.05 (s, CH₃) and 1.09 (s, CH₃) due to

TABLE 1. ¹³C-NMR CHEMICAL SHIFTS OF AGLYCONE MOIETY OF COMPOUND 1 IN CD₃OD.

Carbon	Shift	Carbon	Shift	Carbon	Shift
1	39.6	11	24.5	21	35.0
2	27.0	12	123.5	22	34.0
3	90.5	13	145.37	23	28.5
4	40.0	14	42.6	24	17.0
5	57.1	15	28.9	25	15.9
6	19.3	16	24.93	26	17.8
7	33.9	17	47.6	27	26.4
8	37.5	18	39.6	28	180.8
9	48.7	19	47.39	29	33.5
10	38.1	20	-	30	24.0

TABLE 2. ¹³C-NMR CHEMICAL SHIFT OF α-L-ARABINOPYRANOSYL AND β-D-GLUCOPYRANOSYL.

Carbon	(1"→2') in C ₅ D ₅ N		(1"→3') in MeOH	(1"→4') in C ₅ D ₅ N
	Chemical shift	Chemical shift	Chemical shift	Chemical shift
α-L-Arabinopyranosyl				
1'	104.7	107.0	106.7	106.8
2'	80.6	72.1	73.6	72.8
3'	72.5	83.6	74.6	74.5
4'	68.1	69.5	79.9	69.0
5'	64.7	66.6	66.2	66.1
β-D-Glucopyranosyl				
1"	105.7	106.4	106.3	
2"	75.1	75.7	75.7	
3"	78.3	78.3	78.6	
4"	71.5	71.6	71.4	
5"	78.3	78.8	78.3	
6"	62.5	62.8	62.6	

7 tertiary methyl groups. The anomeric signals at δ4.28 (d, J_{1,2}=7.1 Hz, H-1') and 4.50 (d, J_{1,2}=7.5 Hz, H-1") show diaxial relationship which is consistent for α-L-arabinopyranosyl and β-D-glucopyranosyl respectively and a distorted triplet appeared at δ5.23 (t, H-12) due to endocyclic double bond.

The ¹³C-NMR spectrum of aglycone moiety is given in Table 1 and Table 2 contains a list of ¹³C-NMR signals of β-D-glucopyranosyl and α-L-arabinopyranosyl having (1"→2'), (1"→3') and (1"→4') linkages and has also ¹³C-NMR signals of only α-L-arabinopyranosyl. It is obvious from the chart of Table 2 that when C-1 of glucose is linked with any of the carbon of arabinose then ¹³C-NMR signal of that carbon of arabinose shifts downfield about 8-10 ppm.

When there is (1→2) linkage as in Akebia saponin P_E then C-2' signal is shifted from 72.8 - 80.6 ppm, a 7.8 ppm downfield shift, while Scabioside B isolated from *Patrinia scabifolia* [5] and its ¹³C-NMR signals [6] have [1→4] linkages and the signal of C-4' is shifted from 69.0 - 79.7 ppm, a 10.7 ppm downfield shift. Like this in guaianin N, ¹³C-NMR signal of C-3' is shifted from 74.4 - 83.6 ppm, a 9.2 ppm downfield shift which indicates that C-1" β-D-glucopyranosyl is attached with C-3' of α-L-arabinopyranosyl.

There are 7 methyl peaks in aglycone region of ¹³C-NMR ranging from δ15.9 - 33.5 ppm and a peak of carboxylic acid carbon (C-28) is found at 180.8 ppm. Unsaturated C-12 and C-13 carbons gave signals at 123.5 and 145.37 ppm respectively. A comparison of ¹³C-NMR spectrum of 1 with that of Akebia saponin P_E [4] showed that the chemical shifts due to aglycon moieties in both the saponins are almost the same so the ¹³C-NMR spectrum of the aglycon of compound 1 was established to be oleanolic acid.

The sequence of sugars has been established through FAB mass spectrum (negative mode) which exhibits molecular ion peak at *m/z* 749 (M-H)⁻ corresponds to the formula C₄₁H₆₆O₁₂ of compound 1. The prominent fragment ion peak at *m/z* 587 results from the expected cleavage of glucose moiety i.e. [M-glucose-H]⁻ corresponds to the formula C₃₅H₅₅O₇, and the next prominent ion peak at *m/z* 455 showing cleavage of arabinose i.e. (M-glucose-arabinose-H)⁻ and also corresponds to the formula of aglycon moiety C₃₀H₄₇O₃.

From all these ¹³C-NMR, ¹H-NMR and FAB negative mass spectroscopic evidences mentioned above, the structure of guaianin N was determined as 3-O-(β-D-glucopyranosyl-(1→3)-α-L-arabinopyranosyl)-oleanolic acid. Compound 1 was found to be active against gram negative *Pseudomonas pseudomaliae*.

Brine shrimp toxicity test [1] was also performed. Its LC₅₀ was found to be 621.1715 mg/ml.

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