Pak. j. sci. ind. res., vol. 35, no. 11, November 1992

# GAS CHROMATOGRAPHY OF ORGANOPHOSPHORUS PESTICIDES IN SOILS BY INTERNAL STANDARD METHOD

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(Received June 17, 1991; revised November 14, 1992)

Two types of soils treated with seven organophosphorus pesticides were extracted separately with six solvent systems. The recovery yields from the soils were studied by gas chromatographic technique using an internal standard. The data were subjected to statistical analysis. The best solvent system for best recovery is reported.

Key words: Gas chromatography, Organophosphorus pesticides, Solvent systems.

### Introduction

Organophosphorus pesticides are widely used in agriculture. They show a varied range of chemical stability towards environmental degradation. Sattar [1] critically reviewed the thinlayer chromatography (TLC) analysis of organophosphorus pesticides present in soils, crops, foods, and other environmental samples. Aoki et al. [2] reported the gas liquid chromatography(GLC) method of detecting, some organophosphorus pesticides from rice, wheat, and dried beans with a detection limit from 0.0002 to 0.015 ppm. Cochrane et al. [3] reported a GLC method for detection of pesticides from the apple samples. A few organophosphorus pesticides were analysed by Sattar and Paasivirta [4] from soils by gas chromatography (GC) using an external standard method. Numerous other methods are [5-22] also available for analysing organophosphorus pesticides from soils or other environmental samples. Recently, Sattar and Rahman [23] reviewed a large number of articles on GC of organophosphorus, organochlorine and phenoxy herbicide compounds from various environmental samples. The present investigation, describes a simple and sensitive internal standard GC method for analysing 7 organophosphorus pesticides present in an acidic and a neutral soil conditions.

## **Materials and Methods**

*Pesticides*. Seven commonly used organophosphorus pesticides were used in the study. The general and the chemical names of the compounds are given as follows: (1) Dichlorvos (2,2-dichlorovinyl dimethyl phosphate), (2) Phosdrin (methyl 3-(dimethoxyphosphinyloxy) but-2-enoate; (3)Dimethoate (dimethyl-S-methylcarbamoyl methyl phosphorodithioate); (4) Diazinon (0, 0-diethyl-0-2-isopropyl-6- methylpyrimidin-4-yliphosphothioate); (5) Parathion (0, 0-diethyl -0-4-nitrophenyl phosphorothioate); (6) Malathion (dimethyl-S-1,2-dicarboethoxy ethyl dithiophosphate) and (7) Bromophos (4-bromo-2, 5-dichlorophenyl dimethyl phosphorothionate). The samples (1)–(7) collected

from EPA, USA were reported to be 99 to 100% pure. Phenylphosphonic acid was used as an internal standard. The compounds were prepared as 0.1 % solution in hexane, ethylacetate or dichloromethane.

*Soils*. A noncalcareous dark grey floodplain (neutral) and a red brown terrace (acidic) soils were used in the experiment. The sampling sites were free from usage of any pesticides. The soils were collected at 0-15 cm depth, dried at room temperature and ground to pass through a 10-mesh sieve. Some of the soil characteristics are reported in Table 1.

TABLE 1. SOIL CHARACTERISTICS.					
Characteristics	Noncalcareous dark grey floodplain soil	Red brown terrace soil			
Moisture %	2.0	2.8			
Sand %	22.8	25.8			
Silt %	30.4	58.5			
Clay %	46.8	15.7			
pH	7.3	7.2			
Organic matter %	1.4	1.1			
Soil type	Sandy loam	Clay			

Soil treatment. Five or ten grams of each soil sample was taken in a 200 ml beaker and each sample 200-500  $\mu$ l (for 5g soil) or 500-1000  $\mu$ l (for 10 gm soil) of solutions of all the seven pesticides were added by using micro-litre pipettes, and mixed well with a glass rod. Distilled water 1 or 2 ml was added to each sample to make the soils simulating field conditions and the strirring was continued until the mixtures were uniform. Then 2-3 g of anhydrous sodium sulphate was added to each beaker, again mixed well, and allowed to stand at room temperature (± 32°).

*Extraction*. The samples were extracted separately with the following solvents:

- (i) Ether: chloroform: dichloromethane (1:1:1),
- (ii) Light petroleum ether:benzene: dichloromethane:2propanol (1:1:1:1)

- (iii) Ether: hexane (1:1),
- (iv) Cyclohexane: acetone (7:3),
- (v) Hexane: ether: dichloromethane: 2-propanol (1:1:1:1:),
- (vi) Toluene: cyclohexane: acetic acid (7:2:1).

For this, each sample with 100 flask was treated with 50 ml of one solvent and shaken over Griffin flask shaker for 45 min. The solutions were filtered through filter paper having 2-3 g of anhydrous sodium sulphate. Each residue was extracted further with 50 of the same solvent already used. The extracts were combined, evaporated, transferred to volumetric flasks and the volumes were made upto 10 ml with hexane.

Derivatization and analysis. 250 or 500  $\mu$ l of the soil extracts were transferred to 10 ml glass-stoppered tubes with 25  $\mu$ l of internal standard (sample No.8) solution, 1 ml of methanol and 2-3 ml of diazomethane ether (to give a permanent yellow colour), mixed well and allowed to stand for

TABLE 2. AVERAGE RELATIVE POSITIONS OF GAS CHROMATO-GRAPHIC PEAKS OF EIGHT ORGANOPHOSPHORUS PESTICIDES WITH STANDARD DEVIATIONS (AVERAGE RESULTS FROM 10

CHROMATOGRAMS).

Comp. No.	Pesticides	Position of peak in chart paper				
	and Int. D standard	Distance from start (cm)	Standard deviation			
1.	Dichlorvos	6.82	0.376			
2.	Phosdrin	10.30	0.536			
3.	Dimethoate	16.70	0.484			
4.	Diazinon	18.4	0.550			
5.	Parathion	20.82	0.312			
6.	Malathion	21.36	0.556			
7.	Bromophos	22.42	0.556			
8.	Phenylphospho acid (Int.standa	nic 9.814 rd)	0.528			

40-50 min. at room temperature. The mixture were evaporated with a stream of nitrogen gas. After appropriate dilutions with hexane, the samples were injected  $(1 \ \mu l)$  to the gas chromatographic equipment. Different suitable concentrations of all the 7 pesticides and the internal standard were mixed and the mixtures were subject to gas chromatographic analysis, and compared with chromatographs of soil samples for calculating percentage recovery of pesticides from samples.

A Perkin-Elmer Sigma 3B gas chromatograph equiped with  $F_1$  detector was used. The 25 m quartz/capillary column was packed with SE-30/Silar-10C. The temperature applied for injector was 250° and that of detector was 270°. A programmed temperature was applied, increasing from 80-220° at the rate of 6°/min.

#### **Results and Discussion**

*Model compounds*. The average positions of 8 compound peaks (No.1-8), standard deviation (S.D.) of the peak distances obtained from 10 chromatograms of the 8 pesticide mixtures were given in Table 2.

The average percentage recovery yields of 7 organophosphorus pesticides (compound No. 1-7) in an acid and a neutral soil samples are given in Table 3. The results varied widely with 2 by the soil samples extracted 6 (I-VI) solvent systems. The solvent No. V gave the best recovery yields of all 7 compounds in both soils, whereas the lowest yield of the 7 compounds were recorded with solvent No. VI. The overall recovery result was higher in neutral (94. 15%) than that of acid type (90.55%) of soil regardless of compounds.

A t-test comparison. The recovery yields of 7 pesticides with I-VI solvent systems were compared by t-test (Table 4.). Most of the result differed significantly within I-VI solvent systems. Maximum variations were recorded when compari-

TABLE 3. AVERAGE PERCENTAGE RECOVERY YIELDS OF SEVEN ORGANOPHOSPHORUS PESTICIDES (1-7), IN SOILS BY INTERNAL STAN-DARD METHOD (EACH RESULT FROM AVERAGE OF FOUR CHROMATOGRAMS) AND THE STANDARD DEVIATIONS (S.D.).

Solvent		1		2	:	3		4	4	5	(	6	7	1
system	%	S.D.	%	S.D.	%	S.D.	%	S.D.	%	S.D.	%	S.D.	%	S.D.
Red brow	n terrace	(acid) se	oil (Bha	luka)										
I	95.0	0.11	95.1	0.05	96.0	0.08	93.0	0.09	93.0	0.11	94.6	0.09	95.7	0.08
П	94.1	0.13	91.2	0.21	92.0	0.14	94.3	0.11	95.3	0.10	94.8	0.15	95.2	0.22
Ш	89.1	0.24	88.1	0.19	91.0	0.23	92.2	0.24	90.0	0.19	89.9	0.23	92.5	0.18
IV	83.4	0.21	83.7	0.23	82.0	0.22	886.1	0.21	85.0	0.20	85.0	0.22	87.7	0.17
V	95.7	0.10	0.1	0.07	97.2	0.05	98.1	0.08	97.5	0.05	95.5	0.07	96.8	0.10
VI	79.1	0.21	80.7	0.17	81.4	0.11	83.4	0.14	85.6	0.11	85.5	0.15	86.0	0.11
Noncalco	areous dan	rk grey fl	oodplai	n (neutral)	soil (Bau, I	Mymensi	ngh)							
Ι	97.4	0.09	99.2	0.08	99.3 `	0.07	95.7	0.05	96.8	0.09	98.9	0.08	99.0	0.06
II	96.4	0.11	94.2	0.09	95.7	0.13	97.8	0.12	98.2	0.12	96.9	0.15	98.5	0.10
Ш	93.3	0.10	94.3	0.10	95.6	0.24	96.6	0.11	94.4	0.08	94.9	0.13	95.0	0.14
IV	88.2	0.17	89.0	0.21	85.0	0.19	91.0	0.21	92.3	0.15	92.3	0.16	94.4	0.12
V	98.8	0.05	98.4	0.06	99.5	0.03	98.7	0.03	98.8	0.07	99.1	0.04	99.3	0.05
VI	80.3	0.15	83.0	0.19	85.3	0.20	86.7	0.14	-	0.18	91.2	0.20	89.4	0.16

type	system	Π	ш	IV	v	VI
Acid	I	3.529	12.389	26.232	4.177	22.696
clay	II	-	8.859	21.770	5.339	19.903
soil	Ш		-	13.669	11.819	13.596
	IV	alean-		S 12 P M	21.505	2.847
	V	Strend South	1.1	요즘 같아요.	100.00	20.851
Neutral	A at I	3.216	8.977	13.240	4.303	12.686
sandy	II	data se	5.838	11.036	7.879	13.893
soil	Ш	markin -	- 1	6.634	13.821	10.323
	IV	a Auro	e - 2	5000	16.138	4.277
	V	-	-		-	17.884

 TABLE 4. T-TEST COMPARISON OF THE RESULTS WITHIN

 SOLVENT SYSTEMS FROM I-VI (DEGREES OF FREEDOM:6).

sons were done within the following sont mixtures: I-IV, I-VI, II-VI, II-VI, IV-V and V-VI.

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