

BIOCHEMICAL ALTERATIONS IN THE FEMALE GENITAL TRACT OF OVARIECTOMIZED RATS TREATED WITH AQUEOUS EXTRACT OF *MORINGA OLEIFERA* LAM

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Biochemical alterations have been observed in the uterus, cervix and vagina of ovariectomized rats in relation to antifertility effect of aqueous extract of *Moringa oleifera* Lam. Its administration has increased glycogen, protein, total cholesterol, acid and alkaline phosphatases in all the organs, however, in vagina the glycogen level decreased significantly. Its combined treatment with estradiol acted synergistically, however, with progesterone it behaved antagonistically.

Key words: *Moringa oleifera* Lam, Biochemical constituents Reproductive organs.

INTRODUCTION

Moringa oleifera Lam., commonly known as Sahajana a member of family Moringaceae, manifests significant antifertility effect in rats [1-4]. Of the various fractions, the aqueous extract of its root evokes significant anti-implantation effect [5] at 200 mg/kg dose when given orally for 7 days after coitus (1-7 *post coitum*). Furthermore extract has also been described for its hormonal properties at different dose schedules [6]. It is known that many plants exhibiting antifertility activity also induce biochemical and physiological alterations in the reproductive organs of adult rats [7,8]. These biochemical fluctuations are usually linked with various carbohydrate, protein and lipid metabolic reactions. The present study has, therefore, been undertaken to evaluate the effect of aqueous extract of *M. oleifera* Lam. on the biochemical constituents of the reproductive organs of the ovariectomized rats.

MATERIALS AND METHODS

Fresh roots of *M. oleifera* Lam. were collected from the Campus of Jiwaji University, Gwalior and was chopped, dried and extracted with water as described earlier [3]. A dose of 200 mg/kg, prepared in gum acacia suspension was administered orally with the help of an intragastric catheter. Estradiol dipropionate (Ovocyclin, CIBA, India) and progesterone (Luteocyclin, CIBA, India) were diluted in olive oil and were administered subcutaneously.

Adult, healthy, virgin female rats of Sprague Dawley strain were selected for the present investigation. These animals were maintained under uniform husbandry conditions and were fed with "Liptom Limited" Gold mohur rat pelleted diet and water *ad libitum*. These were ovariectomized bilaterally under light ether anaesthesia and after a post operative rest of two weeks, animals were randomiz-

ed into eight groups of 6 each and received different treatments in the following manner :

Group No.	Treatment
1. OVX	— Ovariectomized control (vehicle only).
2. OVX + EDP	— OVX + estradiol dipropionate (s.c.) 1 µg/rat/day for 7 days.
3. OVX + P	— OVX + progesterone (s.c.) 1.25 mg/rat/day for 7 days.
4. OVX + E + P	— OVX + EDP + P as in groups 2 and 3.
5. OVX + Ext	— OVX + extract (oral) 200 mg/kg for 7 days.
6. OVX + Ext + EDP	— OVX + extract as in group 6 + EDP as in group 2.
7. OVX + Ext + P	— OVX + extract as in group 6 + P as in group 3.
8. OVX+Ext+EDP+P	— OVX + extract as in group 5+EDP and P as in group 2 and 3.

The animals were sacrificed 48 hrs after the last treatment. The organs *viz.* uterus, cervix and vagina were excised, freed from adhering tissue, blotted and weighed to the nearest 1 mg on a single pan electric balance. Fresh tissue was processed for the estimation of glycogen [9] whereas isotonic buffered homogenate was processed for the estimation of proteins [10], acid and alkaline phosphatase activity [11], total and esterified cholesterol [12]. The results were analysed statistically using students 't' test.

RESULTS AND DISCUSSION

Table 1, 2 and 3 represent the effect of aqueous extract of *M. oleifera* Lam. on the biochemical constitu-

Table 1. Effect of aqueous extract of *M. oleifera* Lam. on various biochemical constituents in the uterus of adult ovariectomized rats. (Values are mean \pm SEM and six animals were used in each set).

Treatment	Wet weight (mg/100g b.w)	Proteins (mg/100mg)	Glycogen (mg/100g)	Acid phosphatase (mg P/100g/h)	Alkaline phosphatase (mg P/100g/h)	Total cholesterol (mg/100g)
OVX Control	45.0 \pm 3.6	10.6 \pm 0.9	40.5 \pm 2.3	86.9 \pm 5.9	296.7 \pm 15.7	0.121 \pm 0.006
OVX+EDP	143.6 \pm 8.6 ^a	11.9 \pm 1.0	74.3 \pm 3.4 ^a	121.1 \pm 6.3 ^a	425.3 \pm 25.4 ^a	0.146 \pm 0.008 ^a
OVX+P	63.8 \pm 3.2 ^a	13.5 \pm 0.7 ^a	45.6 \pm 2.8	201.9 \pm 12.8 ^a	378.7 \pm 16.7 ^a	0.163 \pm 0.008 ^a
OVX+EDP+P	122.5 \pm 7.0 ^b	16.2 \pm 0.9 ^b	60.6 \pm 2.6 ^b	99.7 \pm 6.5	289.8 \pm 15.8 ^b	0.160 \pm 0.008
OVX+Ext	100.0 \pm 6.1 ^a	11.2 \pm 0.8	71.2 \pm 3.4 ^a	108.4 \pm 6.5 ^a	430.5 \pm 22.8 ^a	0.140 \pm 0.007 ^a
OVX+EDP+Ext	169.8 \pm 8.9 ^b	12.6 \pm 1.0	85.7 \pm 3.9 ^b	130.7 \pm 7.5	460.0 \pm 23.0	0.156 \pm 0.007 ^b
OVX+P+Ext	89.8 \pm 6.2 ^c	15.3 \pm 0.6 ^c	65.3 \pm 3.4 ^c	160.6 \pm 8.4 ^c	405.3 \pm 22.1 ^c	0.169 \pm 0.008 ^c
OVX+EDP+P+Ext	134.6 \pm 7.8	16.1 \pm 0.8	70.6 \pm 3.5	120.3 \pm 6.5	325.6 \pm 18.7	0.170 \pm 0.009

Values are statistically significant ($P < 0.05$) when compared to OVX Control (a), EDP Control (b) and P Control (c).

Table 2. Effect of aqueous extract of *M. oleifera* Lam. on various biochemical constituents in the cervix of adult ovariectomized rats. (Values are mean \pm SEM and six animals were used in each set).

Treatment	Wet weight (mg/100g b.w)	Proteins (mg/100mg)	Glycogen (mg/100g)	Acid phosphatase (mg P/100g/h)	Alkaline phosphatase (mg P/100g/h)	Total cholesterol (mg/100g)
OVX Control	16.3 \pm 0.9	12.0 \pm 0.8	32.5 \pm 2.2	130.6 \pm 6.6	190.0 \pm 8.5	0.110 \pm 0.006
OVX + EDP	52.0 \pm 2.1 ^a	16.2 \pm 1.1 ^a	56.3 \pm 3.8 ^a	219.0 \pm 12.0 ^a	519.9 \pm 25.2 ^a	0.128 \pm 0.006 ^a
OVX + P	34.3 \pm 2.6 ^a	15.3 \pm 0.8 ^a	38.1 \pm 2.5	158.3 \pm 7.2 ^a	310.7 \pm 15.2 ^a	0.130 \pm 0.007 ^a
OVX+EDP+P	44.1 \pm 2.5 ^b	19.5 \pm 1.1 ^b	46.1 \pm 2.2 ^b	210.1 \pm 10.9	440.5 \pm 26.3 ^b	0.133 \pm 0.005
OVX+Ext	37.8 \pm 2.1 ^a	15.6 \pm 0.9 ^a	48.5 \pm 2.9 ^a	190.2 \pm 9.7 ^a	490.3 \pm 24.9 ^a	0.115 \pm 0.005
OVX+EDP+Ext	56.8 \pm 2.8	15.9 \pm 0.7	59.0 \pm 2.8	230.7 \pm 12.2	590.0 \pm 25.4 ^b	0.120 \pm 0.006
OVX+P+Ext	40.1 \pm 2.4	18.5 \pm 0.9 ^c	44.3 \pm 2.2 ^c	167.7 \pm 8.6	390.2 \pm 20.7 ^c	0.126 \pm 0.005
OVX+EDP+P+Ext	47.3 \pm 2.8	19.3 \pm 1.0	50.4 \pm 3.5	220.4 \pm 12.1	600.7 \pm 30.9	0.131 \pm 0.005

Values are statistically significant ($P < 0.05$) when compared to OVX control (a), EDP control (b) and P control (c).

Table 3. Effect of aqueous extract of *M. oleifera* Lam. on various biochemical constituents in the vagina of adult ovariectomized rats. (Values are mean \pm SEM and six animals were used in each set).

Treatment	Wet weight (mg/100g b.w)	Proteins (mg/100mg)	Glycogen (mg/100g)	Acid phosphatase (mg P/100g/h)	Alkaline phosphatase (mg P/100g/h)	Total cholesterol (mg/100g)
OVX Control	40.1 \pm 2.4	11.6 \pm 0.8	52.1 \pm 3.4	81.3 \pm 5.5	140.5 \pm 8.4	0.102 \pm 0.006
OVX+EDP	77.1 \pm 3.2 ^a	13.3 \pm 0.8 ^a	31.4 \pm 3.7 ^a	131.4 \pm 6.1 ^a	230.3 \pm 14.6 ^a	0.126 \pm 0.005 ^a
OVX+P	60.3 \pm 3.1 ^a	13.8 \pm 0.6 ^a	48.5 \pm 2.6	108.6 \pm 5.4 ^a	168.4 \pm 8.4 ^a	0.120 \pm 0.006 ^a
OVX+EDP+P	67.5 \pm 3.7 ^b	17.5 \pm 0.9 ^b	41.7 \pm 2.6 ^b	114.0 \pm 5.4 ^b	180.3 \pm 10.4 ^b	0.118 \pm 0.005
OVX+Ext	71.3 \pm 4.5 ^a	12.4 \pm 0.6	27.4 \pm 1.4 ^a	110.0 \pm 6.8 ^a	210.3 \pm 12.5 ^a	0.104 \pm 0.006
OVX+EDP+Ext	90.3 \pm 4.2 ^b	15.4 \pm 0.8 ^b	30.1 \pm 2.0	148.0 \pm 5.8 ^b	280.5 \pm 13.2 ^b	0.130 \pm 0.006
OVX+P+Ext	64.5 \pm 3.5	18.1 \pm 0.9 ^c	42.7 \pm 2.3	110.0 \pm 5.8	178.1 \pm 9.2	0.132 \pm 0.006
OVX+EDP+P+Ext	79.8 \pm 4.4	18.0 \pm 0.9	40.3 \pm 2.5	119.0 \pm 6.7	199.7 \pm 8.9	0.121 \pm 0.005

Values are statistically significant ($P < 0.05$) when compared to OVX Control (a), EDP Control (b) and P Control (c).

ents in the genital tract of ovariectomized rats. Administration of estradiol dipropionate and progesterone *per se* increased the wet weight glycogen contents, protein concentration activity of acid and alkaline phosphatase and the level of total cholesterol in the uterus (Table 1), cervix (Table 2) and vagina (Table 3); however, estrogen exerted a more pronounced effect. Further, vaginal glycogen (Table 3) showed significant depletion after the administration of estradiol dipropionate. In their conjoint treatment progesterone tends to antagonise the estrogen induced gain, however, protein content was significantly increased after their treatment.

Administration of the aqueous extract of *M. oleifera* Lam. *per se* elevated significantly the wet weight and other biochemical constituents in the uterus (Table 1), cervix (Table 2) and vagina (Table 3) of spayed rats, however, the vaginal glycogen contents has been decreased when compared to OVX control. Administration of the extract conjointly with estradiol dipropionate further augmented the effect when compared with EDP *per se*. Concurrent administration of progesterone and extract provoked a marginal increase in the wet weight, glycogen contents, activity of acid and alkaline phosphatases and the level of total cholesterol, however, considerable rise is observed in the protein contents of uterus, cervix and vagina.

Ovarian hormones, estrogen and progesterone are known to alter biochemical constituents of the female reproductive tract through their generalized mechanism [13,14]. Administration of estrogen or progesterone to adult ovariectomized rats is known to elevate significantly the wet weight of the uterus [15-17], cervix [18] and vagina [19] albeit, the estrogen is found to exert more pronounced action. On the basis of this inherent virtue, the estrogenic and antiestrogenic nature of many contraceptive agents has been assessed [20-23]. The present findings are in agreement with the above observations. Further, the aqueous extract of *M. oleifera* Lam. increased the wet weight of the uterus, cervix and vagina of the ovariectomized rats, thereby depicting the estrogenicity of the extracts. Synergistic action with estradiol further corroborate its estrogenic activity [6].

A typical estrogen is known to increase the glycogen contents in the uterus [24-27] and cervix [18] of adult rats and also in immature animals [28,29], however, in vagina the glycogen contents are decreased significantly [3]. Progesterone, an active antiestrogen, plays a controversial role. It has been reported to reduce significantly the glycogen contents in the uterus of adult rats [18,25] but simultaneously other authors [24,31,32] have reported no change.

An increase in the uterine acid phosphatase activity has been reported in the immature rats and adult spayed rats after estrogen [16,18,33] and progesterone therapy [34,35]. However, no significant change has also been reported in acid phosphatase activity in the uterus of ovariectomized rats treated with progesterone [36]. Similarly, an increase in uterine alkaline phosphatase activity in both intact and ovariectomized mice and rats after estrogen therapy have been reported by many workers [16,26,29,33]. Progesterone is known to increase [36] or inhibit [34,37,38] or even provoke no change [29,39] in the activity of alkaline phosphatase. It has been reported that the level of uterine cholesterol is not much changed during different phases of the oestrous cycle, ovariectomy and also by steroidal treatment [40]. Rosenman *et al.* [41] have reported a decreased cholesterol synthesis as a result of estrogen treatment. Further Moskowitz *et al.* [42] reported that after 6 weeks of estradiol 17 β treatment, rat showed increased cholesterol.

Our present findings reveal that the administration of aqueous extract of *M. oleifera* Lam. to ovariectomized rats elevated the wet weight, glycogen content, protein concentration, activity of acid and alkaline phosphatase in the uterus, cervix and vagina although the glycogen contents in vagina were decreased and level of total cholesterol showed marginal increase. These findings clearly reveal estrogenic mechanism of aqueous extract of *M. oleifera*. Additionally its synergistic action with estradiol further strengthens the estrogenic property which has already been observed in immature ovariectomized rats [6].

The increase in the uterine and cervical glycogen contents after the treatment with the extract may be due to the increase in the input of more estrogen which consequently increase the glycogenetic process of the carbohydrate metabolism as it is known that estrogenic substances increase the rats of hexokinase reaction [43,44]. The decrease in the vaginal glycogen contents after the treatment with extracts say that in this tissue the estrogen increases the glycogen consumption rate than its storage. The increased uterine glycogen due to estrogenic nature of the aqueous extract of *M. oleifera* Lam. may also be involved in the rhythmic uterine contractions [45] which may further propel the blastocysts from the uterus and thus provoke their anti-implantation action. Therefore, the increased uterine glycogen level as observed in the present study may be involved in providing the ready made energy for the uterine contractions and thus helps in the expulsion of the fertilized eggs from the uterus. This explanation is further strengthened by the fact that the administration of exogenous glycogen in the uterus of pregnant rats terminates

the pregnancy [46]. Increased activity of the acid and alkaline phosphatase in the genital tract under the influence of aqueous extract of *M. oleifera* may be involved in removing the debris or lysed cells [47,48] or it may alter the permeability of the cell membrane resulting in the increased absorption of nutrient material by the cell [49].

Therefore, on the basis of our present study it may be concluded that the aqueous extract of *M. oleifera* Lam. provoked significant alterations in the genital tract probably due to its estrogenic nature. It is expected that due to the disturbances in these biochemical transformations, the extract may create unfavourable conditions in the uterus which may become the cause of infertility.

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REFERENCES

1. B.S. Aswal, D.S. Bhakuni, A.K. Goel, B.N. Mehrotra and K.C. Mukherjee, *Ind. J. Exp. Biol.*, **22**, 312 (1984).
2. B.N. Dhawan, G.K. Patnaik, R.P. Rastogi, K.K. Singh and J.S. Tondon, *Indian J. Exp. Biol.*, **15**, 208 (1977).
3. A.C. Prakash and R. Mathur, *Ind. J. Exp. Biol.*, **14**, 623 (1976).
4. A.O. Prakash, V. Saxena, S. Shukla and R. Mathur, *Acta Europaea Fertilitatis*, **16**, 59 (1985).
5. S. Shukla, R. Mathur and A.O. Prakash, *International J. Crude Drug Res.*, **26**, 29 (1988a).
6. S. Shukla, R. Mathur and A.O. Prakash, *J. Ethnopharmacol.*, **22**, 55 (1988b).
7. H.S. Chakravarti, *J. Indian Med. Assoc.*, **37**, 322 (1961).
8. A.O. Prakash, S. Shukla, A. Gupta and R. Mathur, *Comp. Physiol. Ecol.*, **11**, 4 (1986).
9. S. Seifter, S. Dayton, B. Novic and E. Muntwyler, *Archs. Biochem.*, **25**, 191 (1950).
10. O.H.N. Lowry, J. Rosenbough, A.L. Farr and R.J. Ramlall, *J. Biol. Chem.*, **193**, 265 (1951).
11. C.H. Fiske and Y. Subbarow, *J. Biol. Chem.*, **66**, 375 (1929).
12. A. Zlatkis, B. Zak and A.J. Boyle, *J. Lab. Clin. Med.*, **41**, 486 (1953).
13. A.V. Nalbandov, *Reproduction in Female Mammals and Birds. In: Reproductive Physiology* (D.B. Taraporewala Sons and Co. Pvt. Ltd., Bombay 1970) A.V. Nalbandov ed., 2nd ed. pp.119.
14. D.C. Turner and J.T. Bagnara, *General Endocrinology* (W.B. Saunders Company, Toppen Company Ltd. Tokyo, Japan, 1975).
15. V.A. Drill, *Oral Contraceptives*, ed. Victor A. Drill, The Blakiston Division (McGraw-Hill Book Company, New York, 1966).
16. J.N. Karkun and F.K. Mehrotra, *Ind. J. Exp. Biol.*, **11**, 7 (1973).
17. L.J. Lerner, *Contraception: Chemical Control of Fertility*, ed. Daniel Lednicer, (Marcel Dekker Inc., N.Y., 1969), pp.161.
18. I.C. Datta, J.N. Karkun and A.B. Kar, *Acta Biol. Med. Germ.*, **20**, 155 (1968).
19. P.K. Mehrotra, *Indian J. Physiol. Pharmacol.*, **20**, 235 (1976).
20. R.A. Edgren and D.W. Calhoun, *Contraception, The Chemical Control of Fertility* (R.A. Edgren, Marcel Dekker Inc. N.Y., 1957), pp.537.
21. S.D. Kholkute and K.N. Udupa, *Ind. J. Exp. Biol.*, **14**, 175 (1976).
22. A. Pakrashi and C. Shah, *Indian J. exp. Biol.*, **15**, 1197 (1977).
23. A.O. Prakash and R. Mathur, *Ind. J. Exp. Biol.*, **15**, 1038 (1977).
24. L.M. Demers, K. Yoshinaga and R.O. Greep, *Biol. Reprod.*, **7**, 297 (1972).
25. A.T. Gregoire, H. Ramsay and A. Adams, *J. Reprod. Fert.*, **14**, 231 (1967).
26. S. Mohla and M.R.N. Prasad, *Acta Endocr. (Kbh)*, **62**, 489 (1969).
27. J.R. Wood, T.R. Wrenn and J. Bitman, *Endocrinology*, **82**, 69 (1968).
28. O. Walaas, *Acta endocr.*, **10**, 175 (1952).
29. J.H. Leatham, *Ann. N.Y. Acad. Sci.*, **75**, 463 (1959).
30. G.S. Tripathi, *Comp. Physiol. Ecol.*, **10**, 187 (1985).
31. E.G. Boettiger, *J. Cell. Comp. Physiol.*, **27**, 9 (1946).
32. W.J. Bo and W.B. Atkinson, *Anat. Rec.*, **113**, 91 (1952).
33. F.A. Dugan, B. Radhakrishnamurty, R.A. Rudman and G.S. Berenson, *J. Endocr.*, **42**, 261 (1968).
34. R. Garcia-Bunnell and D. Brandes, *Am. J. Obstet. Gynec.*, **94**, 1045 (1966).
35. A. Goldberg and H.W. Jones Jr., *Obstet. and Gynecol.*, **7**, 542 (1956).
36. M. Hayashi and W.H. Fishman, *Acta endocr. (Kbh)*, **38**, 107 (1961).
37. W.B. Atkinson and E.T. Engle, *Endocrinology*, **40**, 327 (1947).
38. T.C. West and P. Cervoni, *Amer. J. Physiol.*, **182**, 287 (1955).

39. W.B. Atkinson and H. Elftman, *Endocrinology*, **40**, 30 (1947).
40. A. Goswami, A.B. Kar and S.R. Chowdhury, *J. Reprod. Fertil.*, **6**, 287 (1963).
41. R.H. Rosenman, M. Friedman and S.O. Byers, *Endocrinology*, **51**, 142 (1952).
42. M.S. Moskowitz, A.A. Moskowitz, W.L. Bradford and R.W. Wissler, *A.M.A. Arch. Pathol.*, **61**, 245 (1956).
43. W.S. Bulloughs, *J. Endocrinol.*, **6**, 340 (1950).
44. R. Roskoshi Jr. and D.F. Steiner, *Biochem. Biophys. Acta*, **135**, 717 (1967).
45. M.R.N. Prasad and S.P. Kalra, *Proc. Intern. Conf. Planned Parenthood*, 8th Santiago 413 (*Intern. Planned Parenthood Fed.*, London, (1967).
46. D.J. Anderson and N.J. Alexander, *Biol. Reprod.*, **21**, 1143 (1979).
47. M.J. Clyman, *Fertility Sterility*, **17**, 281 (1966).
48. J.T. Valardo, *Ann. New. York Acad. Sc.*, **75**, 441 (1958).
49. L. Bitenski and S. Cohen, *J. Obstet. Gynec. Brit. C. Wealth*, **72**, 63 (1965).

feature of its own. When the female insect feeds on a tree which has apparently received enough water, as in the monsoon season, the growth is excellent but it gives rise to a generation of larvae which are destined to become winged males. The question now arises how does the species manage to exist. The larvae in its second stage or perhaps earlier changes its sex and becomes bisexual. When such an insect forms its cell, it is special to it and appears like a crown. The usual cell of a full grown female would be round or spheroidal, while the full grown bisexual cell would be octagonal and conceived as crown-shaped.

Fig. 1 shows at the bottom an ordinary crown shaped cell. It has given birth to a generation largely constituted of



Fig. 1. A crown shaped or bisexual cell with its progeny consisting mostly of females soon after the third moult or as young adult females.

Karyopsis, *Lac. Insect. Cultivation*.

INTRODUCTION

In order to cultivate the lac insect it is imperative to know its biology. In some areas in India lac is only collected when a tree is found infected with it to the extent that it would pay the trouble of collection. In as much as there is a market for lac many have actually tried to propagate it. Now here comes an interesting biological fact. In Mysore State lac is cultivated on *Shorea* trees and the insect being *Karyopsis*. But lac is also found naturally infected on *Ficus* *sp.* and some other host plants, but in this case the species is *Karyopsis*, but no one ever exploits these trees or better stated propagate the insect. *Karyopsis* strange enough the same species is found in Hyderabad State usually on *F. bengalensis* and *F. religiosa*. Here again no one takes such brood-lac and infect other trees of the same species on which it was found naturally infected. This negligence has been mistaken for want of enterprise on the part of the collector of lac in Hyderabad State, near the town of Nirmal up north near Betul, lac is regularly cultivated on *Bacca* *sp.* and the forest department experts as part of the revenue from lac as minor forest produce. Thus both in Hyderabad and in Mysore State lac is cultivated on some trees while it is only collected and never propagated on other trees. The answer to the above paradox is furnished by the biology of the species *Karyopsis* which is found in both Mysore and in Hyderabad.

I prefer to refer to this species for the *Shorea* lac insect. *Karyopsis*, comes very near to *Karyopsis* in a special