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SYMBIOTIC BIODEGRADATION OF CROP RESIDUES Part I. Biodegradation of Wheat and Rice Straw

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The effect of pre-treatment and reduction in particle size of straws on its microbial digestibility has been investigated. Biodegradation of rice straw was 48.35 % and 69.93 % when *Penicillium* and *B. polymyxa* were employed singly. The combination of *B. polymyxa* with *Penicillium* showed an improvement in the degradability of rice straw upto 71.67 %. The digestibility of cellulose present in wheat straw increased by 54.85 % by symbiotic effect of *B. Cereus* and *Chaetomium*. Reduction in particle size improved the susceptibility to microorganisms.

INTRODUCTION

Crop residues represent one of the largest potential resources for future energy and food. Their nutritive value can be increased by growing cellulase producing fungi on them. Recently a large number of rumen and soil microorganisms have been found which possess the ability to degrade native cellulose into simpler carbohydrates[1-4]. Methods for enzymic hydrolysis of cellulose to glucose especially by the cellulase produced by *Trichoderma* have also been developed [5-9]. Degradation of cellulose has been improved by using mixture of cellulolytic microorganisms than individual enzymes due to their synergistic action[10]. A decrease in particle size increased susceptibility of the substrate to the enzymic hydrolysis because of the increase in surface area[11].

The present study deals with the symbiotic effect of various microorganisms on the degradability of crop residues.

MATERIALS AND METHODS

Wheat and rice straws were purchased from the local market and were ground to 60, 80, 100, 120 and 200 mesh.

Reese medium[12] containing 0.5 % of wheat or rice straw was fermented with molds and bacteria. The fermented residue was centrifuged at 1185g for 10 min. The precipitate was used for the estimation of undigested cellulose and percentage increase in nitrogen.

Analytical methods for the estimation of nitrogen and cellulose were the same as reported elsewhere[13,14].

RESULTS AND DISCUSSION

The Effect of Particle Size on the Growth of Micro-

organisms. Effect of particle size on the biodegradation of wheat straw was first studied with the aim of selecting a mesh size most suitable for the purpose. Accordingly wheat straw was ground from 60 to 200 mesh sizes and biodegradation of the material by *Penicillium*, *Chaetomium*, *Streptomyces*, *Trichoderma* and their combinations was studied. It was noted that particle size has an effect both on the degradation of cellulose and increase in nitrogen content. This effect was most conspicuous from 100-200 mesh.

At 200 mesh, *Chaetomium* degraded cellulose (41.26 %) while a combinations of *Chaetomium* and *Trichoderma* or *Chaetomium* and *Streptomyces* showed maximum degradation (50.29 %).

Increase in nitrogen content was also maximum at 200 mesh. Thus *Trichoderma* alone and a combination of *Streptomyces* and *Trichoderma* increased nitrogen content upto 73.01 %, and 89.68 % respectively.

These observations are in agreement with the findings of other workers[9,11] that by reducing the particle size more surface area is exposed which made the substrate more accessible to the microbial enzymes.

Since grinding of cellulosic material to 200 mesh posed operational problems it was decided to use 100 mesh powder throughout the present studies.

Biodegradation of Straws with Single Strain of Molds, Bacteria and Yeast. The effect of molds, bacteria and yeast on biodegradation of ground straws is shown in Table 1. Maximum biodegradation of cellulose 69.93 % was observed when rice straw was incubated with *Bacillus polymyxa*. Next in line was *Saccharomyces cerevisiae* which degraded 50 % cellulose of rice straw and 35.92 % that of wheat straws. The increase in nitrogen content was 87.06 %. *Penicillium* showed 48.35 % improvement in

cellulose degradation and 152.58 % increase in nitrogen contents. Biodegradation of rice straw and wheat straw with *Saccharomyces cerevisiae* was 50.0 % and 35.92 % respectively. Digestibility of various crop residues with cellulose degrading bacteria was reported by various workers [15, 16, 17].

Symbiotic Biodegradation of Straws with Molds, Bacteria and Yeast. Biodegradation of rice and wheat straws due to combined effect of molds, bacteria and yeast is given in Table 2. It is evident that 47.2 % of the cellulose present in rice straw was biodegraded due to symbiotic effect of *Chaetomium* and *Streptomyces*. The increase in nitrogen contents was upto 147.4 %. The combination of *Chaetomium* and *Penicillium* was also found to be effective in case of wheat straw. The increase in nitrogen in case of wheat straw was 64.51 % which was associated with 44.17 % degradation of cellulose. Similar results were reported by

Peitersen [8] using mixed cultures of cellulolytic fungi.

Symbiosis of *B. polymyxa* with *Streptomyces* degraded 75.85 % of the cellulose present in rice straw with 62.06 % increase in nitrogen content. An increase in nitrogen contents up to 130.17 % and 52.41 % cellulose degradation was observed when *S. cerevisiae* was propagated in combination with *Penicillium* on rice straw. Surender and Gupta [16] reported good growth of *Saccharomyces cerevisiae* on urea enriched hydrolysate of cellulosic material. Peitersen [8] while using *S. cerevisiae* in combination with *T. viride* reported rapid utilization of glucose and production of cellulase. Symbiotic effect of *Penicillium* with *B. cereus*, *B. polymyxa*, *B. sphaericus*, and *B. subtilis* on rice straw resulted in an increased nitrogen contents by 92.24, 70.68, 97.41, 87.06 and 130.07 % respectively.

In wheat straw, maximum biodegradation of cellulose (54.85 %) was observed due to symbiotic effect of *B.*

Table 1. Biodegradation of crop residues by molds and bacteria

Cultures	Rice straw		Wheat straw	
	Degradation of cellulose % age	Increase in nitrogen % age	Degradation of cellulose % age	Increase in nitrogen % age
<i>Penicillium</i>	48.35	152.58	17.99	61.20
<i>Chaetomium</i>	36.36	193.10	26.01	60.48
<i>Streptomyces</i>	34.26	87.06	20.07	42.74
<i>Trichoderma</i>	40.20	172.41	26.69	73.38
<i>Bacillus brevis</i>	43.00	27.58	18.44	31.69
<i>Bacillus cereus</i>	31.46	18.96	12.62	4.83
<i>Bacillus laterosporous</i>	6.29	91.37	13.59	11.29
<i>Bacillus pumilus</i>	34.26	72.41	31.55	8.06
<i>Bacillus polymyxa</i>	69.93	87.06	5.82	8.87
<i>Bacillus sphaericus</i>	33.91	72.41	19.14	16.12
<i>Bacillus subtilis</i>	39.16	60.34	13.59	10.48
<i>Saccharomyces cerevisiae</i>	50.00	18.96	35.92	45.16

Table 2. Symbiotic biodegradation of crop residues by molds bacteria and yeast.

Cultures	Rice straw		Wheat straw	
	Degradation of cellulose % age	Increase in nitrogen % age	Degradation of cellulose % age	Increase in nitrogen % age
A				
<i>Chaetomium + Trichoderma</i>	36.36	123.41	35.04	67.74
<i>Trichoderma + Streptomyces</i>	46.50	131.03	24.03	83.87
<i>Streptomyces + Penicillium</i>	43.70	185.35	20.35	82.25
<i>Penicillium + Chaetomium</i>	46.15	77.58	44.17	64.51
<i>Trichoderma + Penicillium</i>	25.17	82.75	30.87	66.93
<i>Streptomyces + Chaetomium</i>	47.20	147.41	31.84	67.74

table continued

B

	43.00	27.58	18.44	31.69
<i>Bacillus brevis</i> + <i>Chaetomium</i>	49.30	39.79	33.99	4.03
<i>Bacillus brevis</i> + <i>Trichoderma</i>	52.00	44.82	32.03	64.51
<i>Bacillus brevis</i> + <i>Streptomyces</i>	56.99	25.86	9.22	33.06
<i>Bacillus brevis</i> + <i>Penicillium</i>	58.39	40.51	15.53	28.22
	31.46	18.96	12.62	4.83
<i>Bacillus cereus</i> + <i>Chaetomium</i>	38.81	8.62	54.85	107.25
<i>Bacillus cereus</i> + <i>Trichoderma</i>	32.86	72.41	33.00	67.74
<i>Bacillus cereus</i> + <i>Streptomyces</i>	42.30	38.76	33.49	29.03
<i>Bacillus cereus</i> + <i>Penicillium</i>	45.80	92.24	22.33	54.03

C

	6.29	91.37	13.59	11.29
<i>Bacillus laterosporus</i> + <i>Chaetomium</i>	51.04	80.17	18.44	50.00
<i>Bacillus laterosporus</i> + <i>Trichoderma</i>	40.90	81.89	22.33	7.25
<i>Bacillus laterosporus</i> + <i>Streptomyces</i>	34.26	97.41	48.05	61.29
<i>Bacillus laterosporus</i> + <i>Penicillium</i>	39.51	73.27	48.54	34.67
	34.26	72.41	31.35	8.06
<i>Bacillus pumilus</i> + <i>Chaetomium</i>	45.80	124.13	22.33	12.90
<i>Bacillus pumilus</i> + <i>Trichoderma</i>	39.51	81.89	13.10	61.29
<i>Bacillus pumilus</i> + <i>Streptomyces</i>	42.30	80.17	38.34	8.87
<i>Bacillus pumilus</i> + <i>Penicillium</i>	54.54	98.27	48.54	6.45

D

	69.93	87.06	5.82	8.87
<i>Bacillus polymyxa</i> + <i>Chaetomium</i>	57.34	44.82	50.00	64.51
<i>Bacillus polymyxa</i> + <i>Trichoderma</i>	71.67	62.06	26.69	93.57
<i>Bacillus polymyxa</i> + <i>Streptomyces</i>	75.87	62.06	23.30	78.22
<i>Bacillus polymyxa</i> + <i>Penicillium</i>	71.67	70.68	31.55	101.61
	33.91	92.82	19.41	16.12
<i>Bacillus sphaericus</i> + <i>Chaetomium</i>	35.66	76.72	26.21	41.12
<i>Bacillus sphaericus</i> + <i>Trichoderma</i>	33.91	94.82	27.72	46.77
<i>Bacillus sphaericus</i> + <i>Streptomyces</i>	53.14	72.41	41.74	13.70
<i>Bacillus sphaericus</i> + <i>Penicillium</i>	31.46	97.41	10.19	25.00

E

	39.16	60.34	13.59	10.48
<i>Bacillus subtilis</i> + <i>Chaetomium</i>	40.90	1.17	38.34	7.25
<i>Bacillus subtilis</i> + <i>Trichoderma</i>	41.25	72.41	26.25	58.06
<i>Bacillus subtilis</i> + <i>Streptomyces</i>	52.44	51.72	29.12	6.45
<i>Bacillus subtilis</i> + <i>Penicillium</i>	45.10	87.06	26.21	8.87
	50.00	18.96	35.92	45.16
<i>Saccharomyces cerevisiae</i> + <i>Chaetomium</i>	41.25	45.68	47.57	52.41
<i>Saccharomyces cerevisiae</i> + <i>Trichoderma</i>	40.50	53.44	33.00	37.90
<i>Saccharomyces cerevisiae</i> + <i>Streptomyces</i>	46.15	41.37	41.74	51.61
<i>Saccharomyces cerevisiae</i> + <i>Penicillium</i>	52.44	130.17	41.26	24.19

cereus and *Chaetomium*. This was accompanied with 107.25 % increase in nitrogen contents. These results clearly indicated that combinations of *Chaetomium* with *B. brevis*, *B. polymyxa*, *B. subtilis* and *S. cerevisiae* was more effective than the others. These findings are in accord with that of Han, *et al* [10] and Peitersen [8] who reported that bacteria in combination with fungus and yeast utilized more cellulose as compared with fungus alone. Improvement in the biodegradation of cellulose with combinations of bacteria and molds than individual microorganisms is due to the fact that pure culture of single organism had a fixed genetic make up, its physiological capacity and rate of degradation of cellulose is limited but these limitations were overcome by using the combination of bacteria and yeast with molds in the fermentation process.

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