Biological Sciences Section

EXTERNAL MORPHOLOGY OF CHRYSOCORIS STOCKERUS (LINN.) (PENTATOMOIDEA : SCUTELLERIDAE) WITH COMPARATIVE ACCOUNTS ON FOURTEEN RELATED SPECIES FROM PAKISTAN, AZAD KASHMIR AND BANGLADESH AND THEIR BEARING ON CLASSIFICATION

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(Received September 26, 1975; Revised April 14, 1977)

Abstract. The external morphology of *Chrysocoris stockerus* (Linn.) (Scutelleridae) is carried out to understand and evaluate the significance of more reliable characters specially those of metathoracic scent gland ostioles and male and female genitalia and in this light fourteen other scutellerids from Pakistan, Azad Kashmir and Bangladesh representing nine genera, five tribes and four subfamilies are studied. The results are compared with the data available in the existing literature and already existing phylogenetic schemes are briefly reviewed.

The external morphology has long been regarded a conventional tool for taxonomical works and primarily for this reason most of the morphological investigations have been undertaken in the past for providing a better under-standing of these characters. Except that by Fedotov²⁰ not a single species of the present rather economically important family Scutelleridae has been worked out in the literature to date on the pattern of those of Tower,⁸¹ Malouf,⁴⁵ Bonnemaisow,¹⁰ Akba16'7 Beccari and Fenili⁹ and MacGill.⁴⁴

Heteropterous morphology in general and those of scutellerids including also those based on lone species are all reviewed in general works and also in the works dealing with comparative morphology covering one aspect or the other of the Heteroptera. Cobben¹⁴ has cited 352 references; Southwood⁷⁵ 330 references and Southwood and Leston's⁷⁶ 'Land and Water bugs of British isles' has given classified references on all sections.

In the light of above, to review all the morphological accounts in a work of present nature in which emphasis is laid on the aspects of phylogenetic importance would not only be superfluous but also out of place. However, some of the general accounts and particularly those on scutellerids, specially of Vidal,⁸³ Piotrowski,⁶² Leston.³⁴ McDonald,^{49–52} Kumar,^{26–29} and Lattin³² are included in the discussion.

McDonald^{50'51} treated Scutellerinae of the sub-family rank and later⁵² agreeing with Pendergrast ⁶¹ and Kumar^{27'28} raised it to family level but considered it difficult to define the group on the basis of male and female genitalia. Below familial level also, the problem is far from settled. Kumar²⁹ concluded that his studies raised more questions than it answered and thought that their solution rested in an investigation of more members of Pachycorinae and Eurygasterinae. Similarly

McDonald⁵² emphasized the need of further work on the Palaearctic species of Odontoscelini for an adequate definition of the tribe.

It is apparent from above that studies on comparative morphology of scutellerids of Pakistan representing some of the Oriental, Palaearctic and Ethiopian fauna was needed not only for filling the gaps in our knowledge, but also in order to solve many existing phylogenetic problems regarding supergeneric categories within the family and that of the family within the superfamily Pentatomoidea.

Material and Methods

All the specimens used in the present studies were collected by the staff members of the USDA, ARC Research Project A17-ENT-37 from various localities of Pakistan, Azad Kashmir and Bangladesh during expedition programmes. The identifications of the specimens were largely made by Dr. Q. A. Abbasi, former Lecturer, Department of Zoology, University of Karachi and confirmed and compared with types (mostly lodged at British Museum Natural History, London) by the present first author. Some of the identifications were also confirmed by Mrs. G. M. Black and Dr. M. S. K. Ghauri of British Museum Natural History and Commonwealth Institute, London respectively.

The specimens were treated in hot 10% caustic potash (KOH) for obtaining clarity and then were washed with water. For decolorization, chlorination was carried out in potassium chlorate and hydrochloric acid (conc.) (KClO₃ + HCl) in fume cupboard and when the material was completely decolorized, it was washed with water and after passing through the grades of alcohol was stained with acid fuchsin. The material was studied in alcohol and later preserved in a solution of absolute alcohol and glycerine. The mandibular and maxillary stylets with levers were dissected from the head, treated with alcoholic, grades and were mounted in balsam. The genitalia were examined from both dried as well as preserved specimens after treatment. For female genitalia the abdomen was removed from the specimen and boiled in 10% KOH, washed with water and after dissecting the abdomen the spermatheca was removed and studied in glycerine and was then preserved in microvial with a drop of glycerine. The abdomen, then was reattached with the specimen, with the help of insect gum.

In the case of male genitalia after removing the pygophore from the specimen and treating the same with KOH and washing thoroghly with water, was dissected and fully inflated in the same medium in a cavity block and after treating it in different grades of alcohol it was placed on a very fine cotton pad immersed in glycerine in a cavity plate with the help of fine forceps. The aedeagus was examined and later was preserved in microvial with a drop of glycerine.

All the illustrations including those of metathoracic scent gland ostioles, seventh sternum in female, female terminalia, spermatheca, dorsal and ventral views of pygophore, inflated aedeagi and parameres have been drawn to the given scales. The diagrams were made under the Poland stereo-scope and Leitz bionoculars, with various magnifications using squared ocular grid and graph papers. For the male genitalia all the conjunctival appendages were numbered in sequence from the dorsal to ventral surface.

Results and Discussion

External Morphology of Chrysocoris stockerus (Linn.)

Head. Areas and Sulci of the Cranium (Fig. 1). The dorsal view of the adult head capsule of Chrysocoris stockerus (Linn.) (Fig. 1)) is porrected, with prominent laterally projecting compound eyes, and a pair of small shiny ocelli (ocl) in between the eyes. The dorsal cranial sclerites and setae are similar to that of Coridius = (Aspongopus) janus (Fabr.) as reported by Banerji⁸ except that neither occipital sulcus nor its internal ridge is present however the occiput (oc Snodgrass⁷²) is marked behind the compound eyes, and there is a somewhat 'U'-shaped impression between the ocelli which internally bears a complete ridge providing evidence for the presence of a true sulcus.

On the basis of this U-shaped impression the small area enclosed by it may be assumed frons (f). However tentrorial pits are entirely absent as reported by Duporte¹⁷ and Rawat⁶⁷. On the other hand there is no epistomal sulcus between the clypeus and frons, so the region from the 'U'-shaped impression to the attachment of labrum (lbr) with clypeus (cly) may be termed as fronto-clypeal region.

Externally there is nothing to divide the clypeus into anteclypeus (tylus of many authors) and post-clypeus (Matsuda,⁴⁷ MacGill,⁴⁴ Southwood⁷³).

Spooner⁷⁷ states that in Heteroptera the frons and clypeus are always fused. Anteriorly the clypeus slightly bends downward to reach the basal portion of the labrum. The slender, longer, tapering labrum (Fig. 9) is suspended from the clypeus by the intervention of a membrane. Myers⁵⁵ refers it the labrum-epipharynx, while Hamilton,23 Butt,12 MacGill,44 Qadri,64 Qadri and Aziz65 and Akba16 labelled it as labrum. It is chitinized, broad at the base and tapering distally and hardly reaches beyond second labial Ventrally the latter possesses a midsegment. longitudinal line, the labial groove (lbg), which retains the stylet bundle.

Knight,²⁵ Butt,¹² MacGill,⁴⁴ Necomei⁵⁶ and Southwood⁷³ named the mandibular plates as 'Jugum' while Muir and Kershaw⁵⁴ called these as frontal ridge. Spooner⁷⁷ named it as paraclypei but Snodgrass⁷⁰ and Ahmad and Shadab⁴ referred these as 'mandibular plates' and the same term is used presently. These are separated from the clypeus along the clypeal cleft (cc). The pleurostomal suture reported by Banerji⁸ in *Coridius janus* (Fabr.) could not be observed in the present study. Ventral to the mandibular plates (map) are the maxillary plates separated by the mandibulo-maxillary suture (Figs. 2, 'mdms).

Each maxillaryplate (mxp) ventrally produces an exo-skeletal lobe which extends downwards supporting the basal labial segment. These lobes are termed bucculae (Spooner⁷⁷) (Fig. 2, b). Parsons⁵⁸ named these the gular lobes; Crampton and Sprague¹⁵ called these the ventral part of the maxillary plate.

The gula (gu) (Fig. 2) is well developed and there is no mark of identification between the gula, genae and maxillary plates. Genae (ge) are present between the antennae and the compound eyes, posteriorly merging with gula while dorsoanteriorly separated from the antennal base.

Antennae (Fig. 4). The antennae are situated on the sides of the head capsule in front of the compound eyes in antenniferous tubercles (an). Each antenna is surrounded by circum-antennal sulcus (cas) and is five segmented. With the first (basal) segment larger than the second. The latter is the smallest. The third segment is slender and distinctly longer while the fourth segment is longest of all segments and is cylindrical whereas the fifth segment is only smaller than fourth and is somewhat flattened. Both the fourth and the fifth segments appear somewhat medially grooved. The basal portion of first antennal segment is yellowish and the entire antenna is of black colour, and is covered with minute bristles.

Compound Eyes (Fig. 1). The compound eyes are large as compared to the head. Each eye is surrounded by a circumocular sulcus (cos), is sessile and globular in shape. These are very prominent and reddish-orange in colour.

Labium (Figs. 5 & 6). The labium is four segmented (sg1-4), long and slender, reaching third coxae. The segmental nature of labium is important in classification and identification. According to Muir and Kershaw⁵⁴ entire surface of first labial segment is fused with bucculae. Same is true in the present species.

Mandibular Stylets (Fig. 8). The mandibular stylets (mnst) are elongated but shorter than the maxillary stylets. These are long and hollow in structure. Their extreme tips are serrate. The mandibular lever (mnl) is triangular in shape and is directly attached to the stylet base (mnd) and jugal plate ventrad to the antennal base.

Maxillary Stylets (Fig. 7). The maxillary stylets (mxst) are a pair of long, slender, partly sclerotized and curved structures running on the inner surface of corresponding maxillary plates. Maxillary stylets are broader at the base and gradually tapering at the apices with pointed and saw-like tips. The maxillary lever (mxl) is also triangular in shape.

Thorax

Prothorax. (Figs. 10, 11) The prothoracic tergum (Fig. 10) is a large plate covering much of the anterior half of the body. It is more or less hexagonal in shape. The antero-lateral margin of the tergum is turned downward in order to meet the pleural region. Internally a transverse ridge (trr) runs along the middle of the pronotal plate and marks the posterior limit of prothoracic coxal cavity (cxc).

There is no demarcation between the notal and pleural areas. Externally a short pleural sulcus (ps, Fig. 11) divides the pleural area into two sclerites; the larger one proximal to the head is the episternum (ep) and a smaller portion lying away from the head is epimeron (em). Internally the pleural sulcus produces a distinct pleural ridge, from the dorsal surface of which is pronounced pleural apophysis (pleural arm of many authors and pleural taken of Larsen^{30,31}) bending inward towards the sternal apophysis or the furca (fur). The furcal bases are connected by a well developed sterna costa (sc).

(Figs. 12, 13 & 14). The meso-Mesothorax thorax constitutes the largest portion of the ptero-thorax. Its dorsal anterior 1/4 area lies beneath the prothoracic notum. Tergum of the mesothorax is made up of two portions; an anterior notum and a post-notum (Fig. 14). The notum is composed of several distinct regions. The prescutal suture is absent. Two small prealar arms (pra) at antero-lateral angles of pronotum suggest that prescutum (psc) is completely fused with scutum, therefore, the former is represented by its ante-rior margin. A 'U'-shaped parapsidal sulcus (pps) runs in the middle of large scutum (sct.). Tower,⁸¹ Taylo1⁸⁰ and Malouf ⁴⁵ erroeneously called the area bound by parapsidal suture, prescutum, as did many subsequent authors such as Larsen^{30'31} and Parsons.⁵⁹ The internal ridge of the parapsidal sulcus is indistinct. The scutum is located medially, laterally and posteriorly to the parapsidal sulcus (pps).

The anterior notal wing process (anp) arises as a small projection along the lateral border of the scutum, but the posterior notal wing process is not visible dorsally. Posteriorly the scutum ex-

tends onto the scuto-scutellar sulcus (sss) which separates it from the huge scutellum (sctl); the key character of the family. The scuto-scutellar suture is rarely found in Heteroptera. The ridge of scuto-scutellar suture is quite distinct internally. Larsen³⁰ has reported this suture in *Eurygaster* and *Odontoscelis*.

The scutellum (sctl) is a large 'U'-shaped sclerite which projects backwards covering mesothoracic post-notum, metathoracic tergum and all the abdominal tergites completely. The postnotum (pnt) is in the form of a narrow thin and membranous structure Jying beneath the scutellum.

The mesosternum and metasternum are completed fused (Figs. 12 & 15) and bound medially by mesosternum which is separated by sterna-costal suture into the anterior basi sternum (bst) and posterior sternacosta (sc). The sternacosta connects the bases of the mesofurca (Fig. 13, mf). A median longitudinal groove runs through the

basisternum which broadens 'posteriorly.

The mesofurca (Fig. 13 mf.) is T-shaped, a three branched structure, a posterior, a latero-medial and an antero-dorsal branch. The latter carries the thread-like furcal arm (fam) and its terminal funnelshaped apodeme (tap). Malouf⁴⁵ misinterpreted sternacosta in *N. viridula* (L.) as antecostal ridge. The anterior margin of mesocoxal cavity (cxcl) is formed by precoxal bridge which is fused with episternum (ep 1). The pleural ridge (Fig. 13 pr) is reduced, externally represented only by dorsal part of the pleural sulcus (ps) and the pleural wing process. The pleural arms arise near the dorsal end of the ridge. The region anterior to the pleural ridge can be recognized as episternum only. The region posterior to the pleural ridge is greatly reduced and represents epimeron (em 1) which forms a narrow postcoxal bridge. A ventral evagination of the mesopleuron forms a supracoxal lobe (scl). The trochantin beneath the supracoxal lobe is easily removable.

The presternum is indistinctly fused with the meso-sternum. A thick band at the anterior margin (Fig. 13, pp) is a secondary evagination which is continuous with the prepectus of the mesopleuron (Matsuda⁴⁸). Khanna²⁴ and Gupta²² errone-ously called this region the presternum.

Metathorax (Fig. 15). The metathoracic tergum is considerably shorter than the tergum of the mesothorax. The prescutum (psc) is completely fused with the scutum. A broad 'U'-shaped parapsidal sulcus runs in the latter. The scutellum (sctl) is a very thin and narrow plate behind the scuto-scutellar sulcus (sss) and the postnotum (pnt) is visible only laterally below axillary cords of the hind wings.

In metathorax (Figs. 12, 13), the metasternum and metapleuron are completely fused without any trace of the pleuro-sternal suture. The precoxal bridge is divided by the scent groove (or paracoxal suture; Matsuda⁴⁸) into an anterior 'laterale' (pre-episternum and an-episternum) .(Fig. 13, al) and a posterior 'antecoxale' (ketepisternum Fig. 13, anc). The latter is fused with the anterior margin of the metacoxal cavity (cxc2) except at outer end. The trochantin beneath the supracoxal lobe (scl) is easily removable.

The pleural ridge (pr) is vestigeal, externally represented by only dorsal part of the pleural suture and by the pleural wing process. The pleural arm is absent. The furcasternum (frt) extends anterolaterally, anteriad to the inner margin of the meta-

coxal cavity. The meta-furcae (mtf) arise at this point and are smaller than the mesofurcae.

The extended antero-lateral angles of the fu_1 casternum together with the meta-furcae form the inner orifice of the scent vestibule. The vestibule (vet) is an evagination of the anterior laterale which makes a secondary connection with the antecoxale, enclosing the entire scent groove; except its outer end which forms the ostiole (o). The ostiole leads into a narrow peritreme (per) which runs laterally and then anteriorly. The metapleuron anterior, posterior, and outer to the peritreme is rugulose and appears distinct from rest of the meta-pleuron, and forms the evaporatoria (ev.). The structure of peritreme, the ostiole and the evaporatoria is usually consistent within a group and offers important taxonomical characters.

Wings

Forewings (Fig. 14, wng & Fig. 16). Hemelytra or mesothoracic wings are basally coraceous but smooth and membranous apically. The dorsal surface is divided into two parts by a transverse semicircular furrow (scf). The coraceous portion of the forewings is divided into three parts.

The apical membrane (mb) contains a large number of small vein-lets (vlt), which run on the longitudinal axis of the wings. These are simple, unbranched and are arranged into two groups, both having seven or eight veinlets respectively. The outer area of the coraceous basal portion lying the costal margin is the embolium (emb) and is marked off from the rest of the coraceous portion by means of R + M.

Next to the embolium is the corium (co) the largest median, triangular area, which is bound anteriorly by radiomedian vein (R + M) and posteriorly by a claval furrow (cf). Its outer margin is marked off from the apical membrane by the furrow (scf). Area inwardly located to the claval furrow is the clavus (cvl). This portion of forewing is structurally similar to corium and embolium.

Hindwings (Fig. 17). Hindwings appear to be entirely membranous yet the apical portion of the wing is much thicker than the posterior much delicate portion. The hind-wings also have the same margins and angles as the fore-wings.

The anterior margins of hindwings are formed of costo-subcostal vein (C+Sc); next to this vein is radius (R), next to the radius is the median (M) which runs in the longitudinal axis of hindwing.

which runs in the longitudinal axis of hindwing. There are three anal veins ending near the middle lobe of the wing. The second anal vein ends at the third lobe of the wing and the last anal vein is present in the middle of the third lobe of the wing. In the hind wings there are also present two wing folds. The first fold (f1) runs parallel with the margin of the cubitus (Cu) and the second fold (f2) runs parallel with the first anal vein. The terminologies of the wing veins are discussed by Leston.⁴² In resting position the wings are folded at these impressions.

Legs

Prothoracic Legs (Fig. 18). The coxa (cox) is more or less cylindrical in shape and projects out farther from the body than the coxae of the second and third pairs of legs. A basicostal suture (bcs) closely surrounds its proximal end. Posteriorly this suture is very faint, producing a basi-costal ridge internally. Each tibia(tib) is flattened dorso-ventrally and is provided with a powerful spine (tibs) at its distal end.

There are three tarsal segments (tar); first two segments are with distal tarsal spines (trs). The distal tarsal segment is joined with pretarsus (Snodgrass⁷⁰) (post-tarsus of Fox and Fox²¹) which consists of two larger claws (cls), a ventral triangular unguitractor plate (ugt) and the sac-like pulvilli ('pul). Unguitractor spines noted by Parsons⁶⁰ in Saldula spp. and by Akbar⁶ in Leptocorisa spp. are not found in the present species of Chrysocoris.

Mesothoracic Legs (Fig. 19). Unlike the prothoracic, mesothoracic legs articulate at two points so the movement is restricted. The mesothoracic coxae are closer to the body than those of prothoracic coxae. The basicostal suture is distinct and separates it from a very narrow basicoxite. The joints between the mesothoracic leg parts are similar to those of prothoracic legs and the locomotion is described by Lauck³³.

Metathoracic Legs (Fig. 20). Coxae are similar to pro-and mesothoracic ones. Only there is some variation in shape and size of different parts.

Abdomen

Pre-genital Tergites (Figs. 23 & 24). Just behind the postnotum of metathorax, there extends posteriorly the intersegmental membrane which is connected with the first abdominal tergite.

This is membranous and is completely fused with metathorax; it cannot be identified separately. The second abdominal tergite is also reduced and modified. It is sclerotized with a transverse ridge in the narrow middle region. The third tergite is comparatively broader than the second, and is finely united with second tergite.

The posterior boundary of the second tergite is overlapping with the anterior boundary of the third tergite, and the intersegmental membrane can be seen only after moving the overlapping tergites. The fourth tergum is longest of all, and is equal to the length of the 2nd and the 3rd segments combined. The paired impressions of the larval dorsal abdominal scent glands are present between the 3rd and 4th, 4th and fifth and fifth and sixth tergites. The fifth, sixth and seventh tergites are partially fused with one another. The connexivum (conx) is completely segmented and ends at seventh tergite. The inner latero-tergites between the abdominal tergites and connexivum are also completely segmented.

The Pre-genital Sternites (Figs. 21 and 25). The pregenital sternites are almost similar in both sexes. The first sternum has undergone complete fusion with metathoracic sternum and therefore the second sternum represents the first visible ventral sternite. Its anterior margin is concave, anterolateral margins are rounded and posterior margin is convex. The third sternite is a rectangular plate. The fourth sternum is larger than the fifth. The fifth, sixth and seventh sternites like preceeding segments have anterior concave and posterior convex margins. Each sternal sclerites on its peripheral area bears a dark pigmented spot which is divided into two by a transverse impression. Above the transverse impression lies the spiracle (sp) on either side and below it a pair of trichobothria (tb) are placed.

The Genital and Post-Genital Regions (Figs. 26 and 28). The genital region comprises the organs of copulation and oviposition. It consists of ninth segment in the male and eighth and ninth segments in the females. The eighth segment in the male is much modified into a cup-like structure which appears semi-membranous and is uniform in texture (Fig. 22).

Female Genital Region or Ovipositor (Fig. 28). The term ovipositor is used usually in those Heteroptera which have elongate lacineate gonocoxae and gonapophyses. These in the present species are modified into plate-like structures and are, therefore, known as genital plates, as in Pyrrhocoroidea and Coreoidea of the Trichophora. Schaefer,⁶⁸ however, prefers the term ovipositor for both types of structures for both are homologous and are involved in deposition of eggs.

In the female genitalia the eighth paratergites (8th pt) and the first gonocoxae (1st gcx) are the components of the eighth abdominal segment. The paratergites are small and triangular in form and each bears a spiracle (8th spr.) and both being medially fused. Ninth paratergites similarly are small and are quadrangular in form. The ninth paratergites (9th pt) in contrast appear externally the only remnants of the ninth segment. Unlike the eight these are not medially fused and underneath, with the help of a membrane are connected with the post-genital segments. The tenth and eleventh appear quadrangular and are called the proctiger (prr). Latter distally bears the anal opening.

Each first pair of gonapophyses (vesicles of the eighth), the second pair of gonocoxae and the second pair of gonapophyses (vesicles of the ninth) appear medially fused and are partly membranous and semi-sclerotized or sclerotized. Schaefer⁶⁸ has homologised the fused first gonapophyses with the triangulin (tr) and the fused second gonapophyses with the arcus of the earlier authors. However, he did not refer the more posteriorly fused second gonocoxae in his paper. As considered by Dupuis^{18'19}, Stys⁷⁹ and Schaefer⁶⁸ the free part of the gonangulum probably has been lost.

The semi-sclerotized or somewhat membranous anterior extension of the ovipositor termed by Scudder⁶⁹ the genital chamber or "gynatrium" by Stys⁷⁸ is much reduced. A sclerotized saccular structure is connected through a proximal duct at the lateral side of common oviduct. The former appears more expanded and is the spermatheca (Fig. 29) serving for the storage of sperm either received directly from the penis during copulation or as conceived by Scuddei⁶⁹ following Leston³⁴ indirectly by the migration of sperms after copulation from the genital chamber specially in the species of some Scutelleridae.

The spermatheca has the usual pentatomoid form with a bulb (spb), a pump region (spp) with distal and proximal flanges (ds. f; px. f) and a distal spermathecal duct (ds. sp. d). This duct leads to the median dilation (sp.d.) in which median duct (sm. sp. d.) and sclerotized rod are clearly visible.

In some other species of the tribe Scutellerini, however, instead, externally the dilation does not show either median duct or sclerotized rod but, as noted by Pendergrast⁶¹ and later by Kumar²⁹, it possesses ornamental lines and appears heavily sclerotized. The proximal spermathecal duct (px. sp. d) first dilates (px. s. sp. d) and then leads to the female's gonopore. Holes in the spermatheca are not observed as noted by Leston³⁴ and Scuddei⁶⁹ which according to them in *Hotea* spp. are produced by insertion of the aedeagus during copulation.

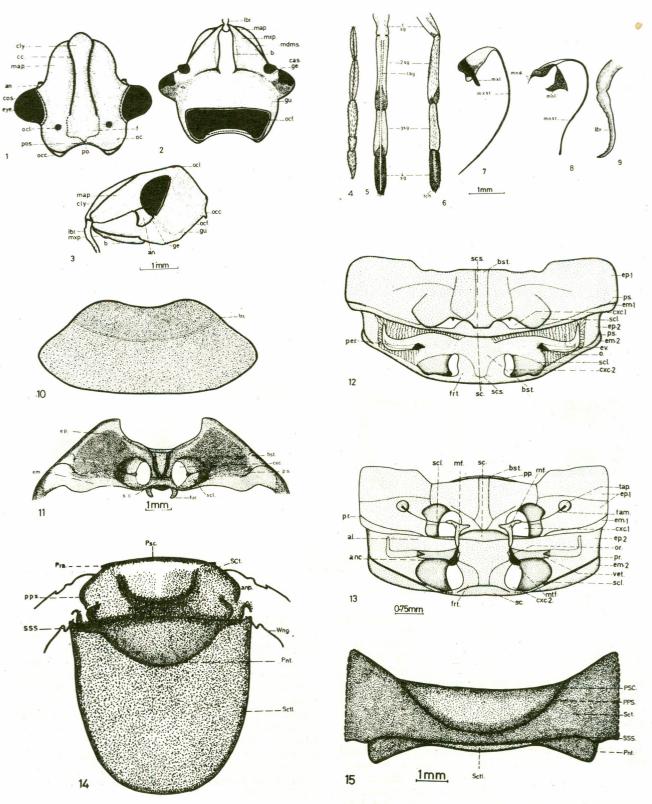
Male Genitalia (Figs. 26, 27, 30, 31, 32 & 33). In Tuxen⁸² Dupuis and Carvalho have given the pattern of the heteropterous genitalia. Ahmad and Southwood⁵ have given hypothetical diagram of male aedeagus labelling all the thecal and conjunctival appendages following their position.

In the male genitalia the ninth segment is modified into characteristic cup-shaped structure known as the pygophore (Figs. 26 and 27, p) in which the organs of copulation are retained. The pygophore has two openings ; the anterior or proximal one an the dorso-posterior, distal, or terminal one. Througd the anterior opening it is connected with the eighth segment which is also modified and membranouh in nature and acts as a connecting membranes between the seventh segment and male genital capsule. The dorso-posterior opening is larger and external in appearance and during copulation the aedeagus with its appendages is inflated through this opening (Bonhag & Wick¹¹, Qureshi *et al.* ⁶⁵). The fused tenth and eleventh segments are in the form of a quadrangular structure, the proctiger (Fig. 26, prr).

(Fig. 26, prr). The dorsal margin of pygophore appears arched and is turned anteriad and appears raised. In some other species on its inner sides processes are seen and these also appear of taxonomic importance. Kumar²⁹ has found processes longitudinal and transverse on the main body of the pygophore ventrally which have not been observed in the present studies.

Within the pygophore on either side of the aedeagus proximally connected with the basal plate

EXTERNAL MORPHOLOGY OF Chrysocoris stockerus (LINN.)



Figs. 1—15. Chrysocoris stockerus (Linn.). 1. Head, dorsal view, 2. Same, ventral view, 3. Same, lateral view, 4. Antennae, dorsal view, 5. Labium, ventral view, 6. Same, lateral view, 7. Maxillary stylets, lateral view, 8. Mandibular stylets, lateral view, 9. Labrum, lateral view, 10. Prothorax, dorsal view, 11. Same ventral view, 12. Mesothorax, dorsal view, 13. Same ventral view, 14. Scutellum, dorsal view, 15. Metathorax, dorsal view,

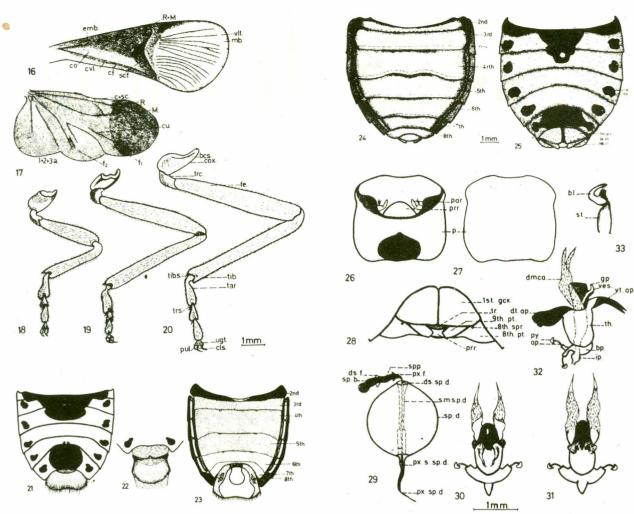


Fig. 16–33. Chrysocoris stockerus (Linn.). 16. Fore-wing, dorsal view, 17. Hind-wing, dorsal view, 18. Fore-leg, lateral view, 19. Middle-leg, lateral view, 20. Hind-leg, lateral view, 21. Male abdomen, dorsal view, 22. Same, showing 8th abdominal segment, dorsal view, 23. Same, ventral view, 24. Female abdomen, dorsal view, 25. Same, ventral view, 26. Pygophore, dorsal view, 27. Same, ventral view, 28. Female terminalia, ventral view, 29. Spermatheca, dorsal view, 30. Aedeagus, dorsal view, 31. Same, ventral view, 32 lateral/view, 33. Paramete, inner view.

are a pair of pointed structures known as parameres (Fig. 26, par; Fig. 23), herpagones, gonostylus or the claspers (Ahmad and Southwood⁵) which probably help in copulation and grasping (Fig. 26, par). Each paramere (Fig. 33) consists of a long, broad stem (st) which bears a curved blade (bl) at the distal end. The proximal inner edge of the blade is beset with dense setae. Aedeagus (the fused mesomeae of Snodgrass⁷¹) is the main copulatory organ (Figs. 30, 31 and 32). It is composed of a proximal basal plate (bp) with pivots (py) or apodemes (ap) and distal phallosoma or theca (th). The latter similar to many alydids (Ahmad and Southwood⁵) bears dorsal (dt. ap) and ventral (vt. ap) sclerotized appendages or processes (also noted and/or drawn by Pruthl63, Kumar^{26^{·27'29}} and McDonald⁵² without comment) Ahmad and Southwood⁵ found these appendages as of great taxonomic significance in Alydidae and probably these are of great taxonomic im-(Kuma1 26'27'29. Scutelleridae portance in McDonald^{50'52} and Lattin³²).

Within the phallosoma or theca lies the conjunctiva or the endosoma of Pruthi⁶³. In the inflated condition it usually possesses three pairs of appendages *i.e.*, the dorsal membranous conjunctival appendages (dmca), the dorsolateral conjunctival appendages (dmca), the dorsolateral conjunctival appendages (Ahmad and Southwood⁵); McDonald's⁵² terminology of first, second and third refers to dorsal, dorso-lateral and frontal in the present sequence; however, these could be confusing when the above author has confused these appendages with those of phallosomal processes (McDonald⁵²). On the other hand Kumar²⁹ has reversed the sequence of McDonald⁵².

The ejaculatory reservoir (McDonalc^{50'52} and the conducting chamber (Kumai^{28'29}) is remarkably developed in this species. Ventral to which is attached the socalled vesica (ves). This is entirely sclerotized, short and curved dorsally and bears the gonopore (gp). McDonald^{49 50'52} has used the structure as taxonomic tool and has

found it usually well developed in the entire tribe Scutellerini.

Kumar ^{28,29} has called this structure the conducting chamber having conducting canals and, has attributed the function of controlling the flow of sperm through gonopore by this chamber and has described a very complicated structure with double system of canals. Previous authors including McDonald ^{49,50,52} have attributed the function of storage of sperms by this region. The histology of this region might further throw some light on the functional aspects of this organ.

The ejaculatory duct before entering into the basal plate forms a funnel-like vesicle forming an inflatory pump (ip) which pushes the semen into the aedeagus during copulation. In the present species as well as in many other scutellerines part of the ductus ejaculatorius and inflatory vesicles are strongly sclerotized and appear-like proximal processes of aedeagus. At the antero-lateral angles of the basal plate the dorsal knobs are produced into apodemes (ap) on each side which bear a pivot (py) at their distal end (Marks⁴⁶), but these do not appear to have taxonomic significance.

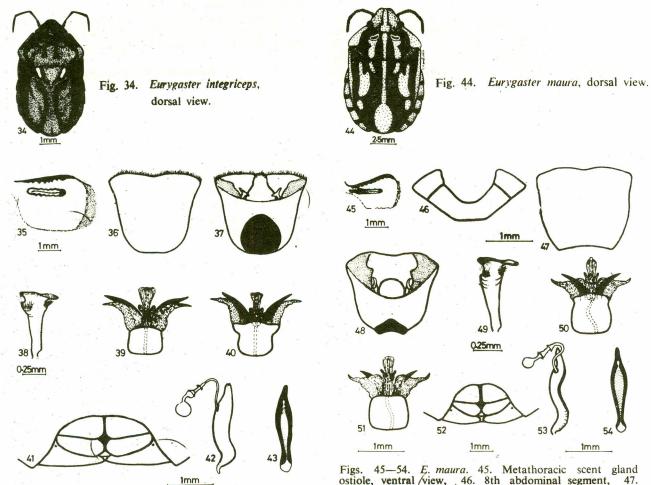
Comparative Morphology

Table 1 gives a list of 15 species of the family Scutelleridae, presently studied with their tribal and sub-familial categories, with figure numbers referring to the illustrations through out the text and relevant descriptions, if any on any of the above categories in the literature todate.

Tables 2-5 give comparative accounts on head, metathoracic scent gland astioles, male and female genitalia and comparisons with other scutellerines reported in the literature are made under phylogenetic considerations.

Species examined	Figs. cited in the text	Previous descriptions
Subfamily: Eurygastrinae		
Tribe: Eurygastrini		
1. Eurygaster integriceps 2. Eurygaster maura	(34-43) (44-54)	Kumar ^{28,29} (conducting chamber, male and female genitalia, Fedetov, ²⁰ Lattin, ³² McDonald ⁵² (male and female genitalia), Lodos ⁴³ (only female terminalia), Piotrowski, ⁶² Vidal ⁸³ and Wagner ^{84,85} (on inflated aedeagus), Scud- der ⁶⁹ (only female genitalia).
Subfamily: Odontoscelinae		
Tribe: Odontotarsini		
3. Alphocoris lixoides	(55-63)	Kumar ²⁹ and McDonald ⁵² (only male and female genitalia).
Subfamily: Pachycorinae		
Tribe: Pachycorini		
4. Deroplax sp.	(64-70)	Kumar ²⁸ , ²⁹ (only conducting chamber, male
5. Deroplax zahidae (MS) 6. Hotea curculionoides	(71-74) (75-78)	and female genitalia), Lattin, ³² McDonald ⁴⁹⁻⁵² (male and female genitalia), Leston ³⁴ (only male
7. Hotea nigrorufa	(79-82)	genitalia), Pendergrast ⁶¹ (only spermatheca), Scuddei ⁶⁹ (only female genitalia).
Subfamily: Scutellerinae		
Tribe: Elvisurini		
8. Solenostethium rubropunctatum	(83-93)	Kumar ²⁸ , ²⁹ (male and female genitalja), Leston ³⁴ (male genitalia), McDonald ^{49_51} (male genitalia and female terminalia).
Tribe: Scutellerini		
9. Cantao ocellatus	(94-104)	Kumar ²⁶⁻²⁹ and McDonal d ⁴⁹⁻⁵² (male
10. Chrysocoris stockerus	(1-33 & 105)	and female genitalia), Leston ³⁴ (male genitalia),
11. Poecilocoris latus	(115-124)	Pendergrast ⁶¹ (only spermatheca), Pruthi ⁶³ (only male genitalia), Scuddei ⁶⁹ (female termi-
12. Poecilocoris (Parapoecilocoris) interruptus. (Probably anohter genus)	(140-149)	nalia and spermatheca).
13. Poecilocoris sp.	(106-114)	
14. Scutellera fasciata	(125-135)	
15. Scutellera nobilis	(136-139)	

TABLE 1.



Figs. 35-43. E. integriceps. 35. Metathoracic scent gland ostiole, ventral view, 36. Pygophore, ventral view, 37. Same, dorsal view, 38. Paramere, inner view, 39. Aedeagus dorsal view, 40. Same, ventral view, 41. Female terminalia, ventral view, 42. Spermatheca, dorsal view, 43. Median dilation dorsal view dousal view.

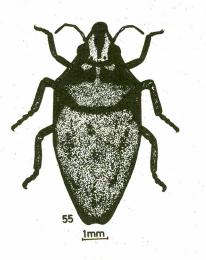
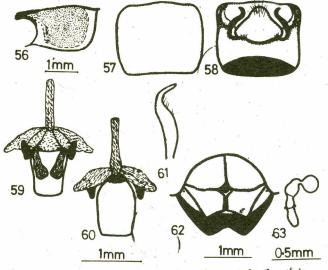


Fig. 55. Alphocoris lixoides, dorsal view.

Imm 0.25mm 53 1mm 1mm

Figs. 45—54. E. maura. 45. Metathoracic scent gland ostiole, ventral /view, 46. 8th abdominal segment, 47. Pygophore, ventral view, 48. Same, dorsal view, 49. Paramere, inner view, 50. Aedeagus, ventral view, 51. Same, dorsal view, 52. Female terminalia, ventral view, 53. Spermatheca dorsal view, 54. Median dilation, dorsal view.



Figs. 56—63. A. lixoides. 56. Metathoracic scent gland ostiole ventral view; 57. Pygophore, ventral view, 58. Same, dorsal view, 59. Aedeagus, ventral view, 60. Same, dorsal view, 61. Paramère, inner view, 62. Female terminalia, ventral view, 63. Spermatheca, dorsal view.

EXTERNAL MORPHOLOGY OF Chrysocoris stockerus (LINN.)

0-25mm

0-5mm

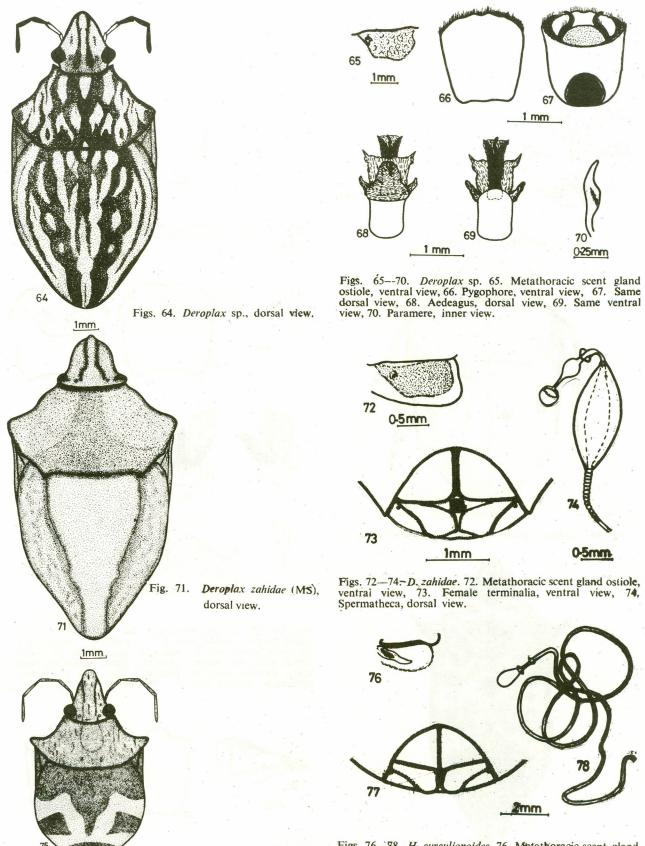


Fig. 75. Hotea curculionoides,

dorsal view.

2mm

Figs. 76—78. H. curculionoides. 76. Metathoracic scent gland ostiole. ventral view, 77. Female terminalia, ventral view, 78. Spermatheca, dorsal view.

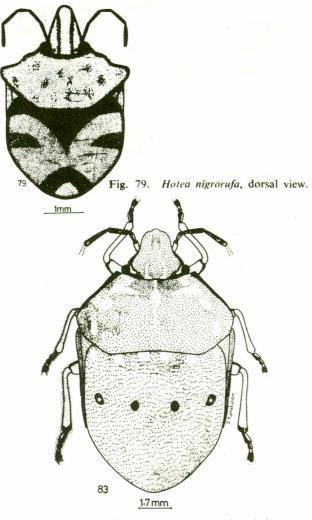
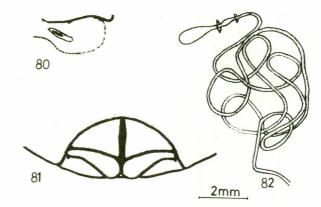


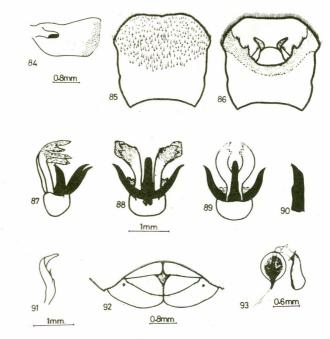
Fig. 83. Solenostethium rubropunctatum, dorsal view.



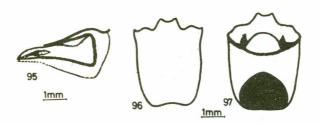
Fig. 94. Cantao ocellatus, dorsal view.



Figs. 80–82, *H. nigrorufa.* 80. Metathoracic scent gland ostiole, ventral view, 81. Female terminalia, ventral view, 82. Spermatheca, dorsal view.



Figs. 84–93. S. rubropunctatum. 84. Metathoracic scent gland ostiole, ventral view, 85. Pygophore, ventral view, 86. Same, dorsal view, 87. Aedeagus, lateral view, 88. Same, dorsal view, 89. Ventral view, 90. Fused vesica and penal lobes, anterior view, 91. Paramere, inner view, 92. Female terminalia, ventral view, 93. Spermatheca, dorsal view.



Figs. 95-97. C. ocellatus. 95. Metathoracic scent gland ostiole, ventral view, 96. Pygophore, ventral view, 97. Same dorsal view.

EXTERNAL MORPHOLOGY OF Chrysocoris stockerus (LINN.)

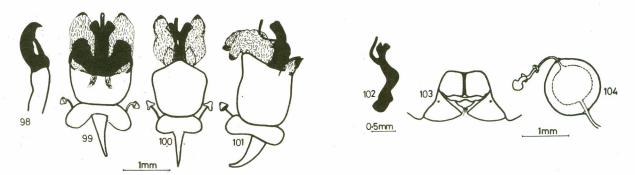
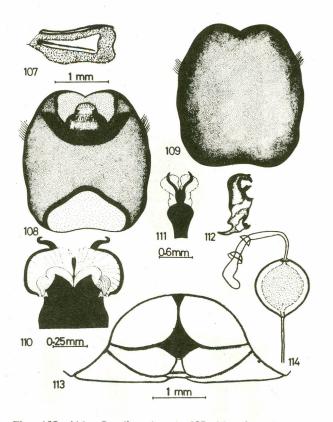


Fig. 98-104. C. bcellatus. 98. Paramere, inner view, 99. Aedeagus, ventral view, 100. Same, dorsal view, 101. Lateral view 102. Fused vesica and penal lobes, anterior view, 103. Female terminalia, ventral view, 104. Spermatheca, dorsal view.



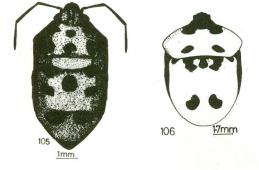
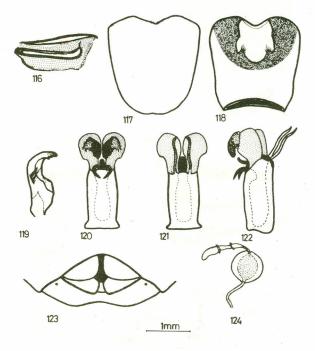


Fig. 105. *Chrysocoris stockerus*, dorsal view.Fig. 106. *Poecilocoris* sp., dorsal view.



Figs. 116–124. *P. latus*. 116. Metathoracic scent gland ostiole, ventral view, 117. Pygophore, ventral view, 118. Same, dorsal view, 119. Paramere inner view, 120. Aedeagus, dorsal view, 121. Same, ventral view, 122. lateral view, 123. Female terminalia, ventral view, 124. Spermatheca, dorsal view.

Figs. 107—114. *Poecilocoris* sp. 107. Metathoracic scent gland ostiole. ventral view, 108. Pygophore, dorsal view, 109. Same, ventral view, 110. Aedeagus, dorsal view, 111. Same ventral view, 112. Paramere, inner view, 113. Female terminalia, ventral view, 114. Spermatheca, dorsal view.



Fig. 115. Poecilocoris, dorsal view.

I. AHMAD and S. MUSHTAQ

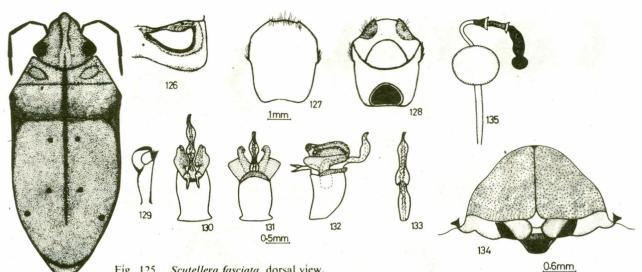
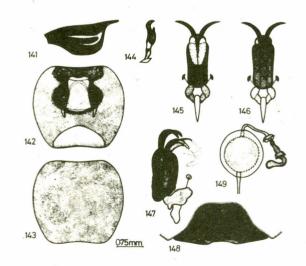


Fig. 125. Scutellera fasciata, dorsal view.

Figs. 126—135. S. fasciata. 126. Metathoracic scent gland ostiole, ventral view, 127. Pygophore, ventral view, 128. Same, dorsal view, 129. Paramere, inner view, 130. Aedeagus, dorsal view, 131. Same, ventral view, 132. lateral view, 133. Fused vesica and penal lobes, anterior view, 134. Female terminalia, ventral view, 135. Spectrate these dorsel view. ventral view, 135. Spermatheca, dorsal view.



Figs. 140. Poecilocoris (Parapoecilocoris) interruptus (probably a distinct genus) dorsal view.



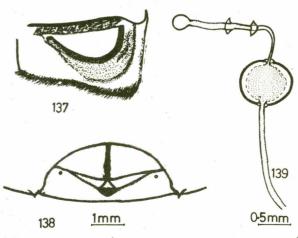
Figs. 141—149. P. (P) *interruptus* (probably a distinct genus). 141. Metathoracic scent gland ostiole, ventral view, 142. Pygophore, dorsal view, 143. Same, ventral view, 144. Para-mere, inner view, 145. Aedeagus, dorsal view, 146. Same, ventral view, 147. Lateral view, 148. Female terminalia, ventral view, 149. Spermatheca, dorsal view.



2mm

125

Fig. 136. Scutellera nobilis, dorsal view.



Figs. 137—139. S. nobilis. 137. Metathoracic scent gland ostiole, ventral view, 138. Female terminalia, ventral view, 139. Spermatheca, dorsal view.

		lypeus		Ge	celli
Species	Length in proportion to mandibular plates	Shape	Mandibular sulci		Inter-ocellar distance in proportion to distance from an ocellus to an eye
1	2	3	4	5	6
E. integriceps	As long as	Convex, raised in the middle.	Very prominent, marked with thin lines.	Small, rounded and some-what raised.	Closer to eyes than to each other.
E. maura	As long as	Convex, deflected anteriad.	Less prominent.	Very small and rounded.	Closer to eyes than to each other.
A. lixoides	As long as	Raised throughout its length.	Less prominent.	Very minute and depressed.	Much closer to eyes
Deroplax sp.	Extending beyond	Convex and raised posteriorly.	Marked with thin lines.	Very prominent and large.	Much closer to eyes.
D. zahidae	Extending beyond	Smoothly convex posteriad.	Less distinct.	Very prominent.	Much closer to eyes.
H. curculionoides	Extending beyond	Anteriorly much raised, convex.	Marked with thin lines.	Very prominent.	Much closer to eyes
H. nigrorufa	Extending much beyond	Anteriorly much raised, convex.	Absent or indistinct.	Very prominent.	Much closer to eyes
S. rubropunctatum	Extending beyond	Convex and raised anteriad.	Somewhat faint.	Very prominent and large	e. Much closer to eyes.
C. ocellatus	Extending beyond	More convex, raised anteriad and posteriad.	Marked with faint groove.	Very large and prominer	closer to eyes.
C. stockerus	Slightly extending beyond	Convex, moderately raised throughout.	Marked with deep grooves.	Moderate but prominen	t. Closer to eyes.
P. latus	Extending far beyond	Broadly convex, raised anteriad.	Marked with deep grooves.	Small, prominent and rounded.	Closer to eyes.
Poecilocoris sp.	Slightly extending beyond	Less convex, raised anteriad.	Marked with deep, anterior and posterior lines, medially very faint.	Large prominent, not raised.	Closer to eyes.
S. fasciata	Extending beyond	Much convex, raised anteriad.	Anteriorly marked with deep groove, posteriorly with thin line.	Large, prominent and rounded.	Closer to eyes.
S. nobilis	Extending beyond	Much convex, raised anteriad and posteriad.	Anteriorly marked with deep groove, posteriorly with thin lines.	Prominent and oval.	Much closer to eyes.
(P.) interruptus Westwood (but probably another genus)	Slightly extending beyond	Broadly convex, raised anteriad and posteriad.	Marked with deep ante- rior groove, posteriorly marked with faint line.	Very small, rounded and raised.	Much closer to eye

TABLE 2. COMPARATIVE CHARACTERS OF HEAD.

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Phylogenetic Considerations

Whereas Leston³⁴–³⁸, China and Miller,¹³ Miyamoto⁵³ and McDonald ^{49°50°51} considered subfamilial status of Scutelleridae as valid, the general concensus has been for the familial status of the group. Pruthi⁶³ though treated the group as a subfamily within the family Pentatomidae but regarded its male genitalia on the same plan as that of Plataspidae and Cydnidae and considered it somewhat more specialized. Southwood⁷⁴ and Nuortera⁵⁷ considered its salivary glands distinct among Pentatomoidea.

Leston³⁹ for the first time gave the group family rank without comment and in the following year⁴⁰ considered Scutellerinae excluded from Pentatomidae. In the same year⁴¹ he considered on the basis of chromosome number that the central group of pentatomoid families represented by the series Tessaratomidae, Scutelleridae and Cydnidae with Phloeidae an early off-shoot of the first named and Brachyplatidae (and probably Aphylidae) an evolved off-shoot of the second. Contrary to Pruthi⁶³ he regarded Scutelleridae constituting the group of lower Pentatomoidea with Acanthosomatidae, Tessaratomidae and Brachyplatidae.

Similarly Cobben¹⁴ discussing the phylogeneses of eclosion in association with egg shape and the micropylar ring in pentatomoids has shown scutellerids somewhat closer to the ancestral form retaining some of their ancient features as also regarded by Ahmad and Abbasi.³

McDonald⁵² while finally recognizing the family status of Scutelleridae found it very difficult to define and distinguish it on the basis of male and female genitalia.

Although Leston⁴¹ and Cobben¹⁴ found the group very stable which is also apparent from the structure of scent gland ostioles and male and female genitalia similar to other externally visible consistent characters like highly developed scutellum (discussed below) the group, nevertheless shows remarkable variations generally in the structure of male and female genitalia.

The evaporatoria in the scent gland ostioles is usually highly developed and is different in texture in comparison to the general pleural surface. In the males the parameres are always with a more or less erectstem and a long (almost an independent looking) or a short (appearing a part of stem) rather curved and lunate blade. The ejaculatory duct before entering into the aedeagus is generally sclerotized. In some species even the inflatory vesicle is also remarkably sclerotized. Theca is always well marked and sclerotized, sometimes provided with dorsal and ventral processes as in scutellerines.

There are usually three pairs of conjunctival appendages. The dorsal membranous conjunctival lobes are usually well developed, sometimes partly sclerotized and partly membranous, occasionally subdivided into 2-3 apical processes. The dorsolateral appendages rarely uniformly sclerotized are usually membranous, sometimes appearing like conjunctival lobes whereas the frontal or ventral pair of appendages except in *Alphocoris* are always membranous, usually very small and unless the aedeagus is fully inflated difficult to be detected.

Penal lobes (of McDonald⁵², Abbasi and Rishi¹ and Ahmad and Abbasi²) a pentatomid character which are developed to support the vesica are entirely absent, instead usually the conducting chamber (Kumar²⁹) or the ejaculatory reservoir (McDonald⁵²) is usually very well developed. In some species of Scutellerini the ejaculatory reservoir itself supports the vesica and in one species (*Cantao ocellatus*) the bilobed remarkably elongated ejaculatory reservoir appears like penal lobes of many pentatomids. It could well be that ejaculatory reservoir developed remarkably in some scutellerids to support vesica before the penal lobes developed in the members of Pentatomidae.

The female genitalia including the spermatheca as suggested by McDonald^{51'52} and later by Schaefer⁶⁸ are on the same pentatomid plan. The spermatheca which appeared different to Pendergrast⁶¹ from other pentatomids, probably only because he studied five species, and out of which four species belonged to a single sub-family Scutellerinae.

No doubt in the majority of the species in the subfamily the spermathecae are characterized by a long tubular apical bulb, a long pump region and a large spherical or ovoid dilation. In some as Pendergrast (op. cit.) has observed, this dilation is heavily sclerotized and deeply pigmented. Not only that, as the above author has observed in *T. diophthalmus* of another subfamily Pachycorinae, this structure is lacking, instead the dilation is thin-walled, is saccular and unsclerotized but also in some other scutellerines (Kumar ²⁶,²⁷ and present studies) thin—walled unsclerotized dilation is present with clearly visible pentatomid-type or sclerotized rod. It could well be that pentatomid type of spermatheca in these species indicate their advanced position in the group.

Below family level the characters of scent gland ostioles and male and female genitalia are more useful in demarcating various supergeneric categories and in showing their mutual relationships.

Eurygastrinae: Leston³⁴ included in his tribe Eurygastrini the subtribes Odontocelaria, Odontotarsaria and Eurygastraria. McDonald⁵² has not only agreed with Lattin³² in separating Eurygastrini from other subgroups of Leston (op. cit.) but also in his final scheme raised the group to subfamily rank. However, he disagreed with the action of Wagner⁸⁵ in raising the group to family status.

The present studies support McDonald's⁵² action. Eurygastrinae with nearly equally developed clypeus and mandibular plates, remarkably broad head, smoothly tapering anteriad with thin slit-like short transverse peritreme of metathoracic scent gland ostioles having wrinkled anterior and posterior margins, and in the males with T-shaped parameres, membranous simple vesica with wide opening and without inner supporting appendages, highly sclerotized dorso-lateral conjunctival appen-

dages and in females with elongated and sclerotized spermathecal dilation clearly stands apart from other subfamilies of Scutelleridae.

In fact the members of this subfamily appear apparently more related, to some of the members of the subfamily Scutellerinae rather than Odontocelinae or Pachycorinae in having distinct peritreme of metathoracic scent gland ostioles and relatively broad head which is much more broader than long.

On the other hand it could well be that the less developed peritreme and other characters of male and female genitalia listed in the Tables 3-5 specially of spermatheca relate the group to some members of Pachycorinae and that the members of this subfamily similar to some of the scutellerines convergently have specialized to suit special habitus and habits.

Distant¹⁶ has included *Melandoema* Jakovlev from Turkestan and North West India in this tribe. Although he has not commented on its metathoracic scent gland ostioles but in his diagram neither the peritreme nor the ostiole are distinct and scutellum is also practically covering the entire abdomen. In view of these two characters it appears more closely related to Odontotarsini (discussed below).

Qdontocelinae: After removal of Eurygastraria, the remaining groups of Leston's³⁴ tribe Eurygastrini are costomarily grouped under Odontocelini to which McDonald⁵² in his final scheme has alloted a subfamilial rank. Distant¹⁶ has treated the two divisions Odontotarsaria and Odontocelaria separately. The present sample is very poor including only one species and therefore it would be unwise to make any conclusive statement at this point.

However, the lack of distinct scent gland ostioles alongwith the peritremes, in the males, the entirely membranous dorsal conjunctival lobes and greatly sclerotized frontal conjunctival appendages with broad rounded apices and in the females very small spermatheca, with indistinct distal flange and median dilation, clearly separate this group from other subfamilies of Scutelleridae. The elongated body, the elongated head, the remarkably developed ninth paratergites and the absence of median dilation in the spermatheca show the group somewhat primitive and in some way relate it to some of the Pachycorinae.

Pachycorinae: During the present studies only representatives of two genera Hotea and Deroplax are included. Kumar²⁹ and McDonald⁵² have included representatives of six and ten genera respectively. The former author has included the presently studied genera in his studies and the latter has included a species of *Tetyra* for which Stål (in Distant¹⁶) raised the group Tetyraria including the presently studied genera Hotea and Deroplax and was followed in this by Distant¹⁶. Kumar²⁹ has found in Hotea and Deroplax one pair of conjunctival processes and a similar type of conducting chamber and on the basis of these characters he has refuted Leston³⁴ who suggested to raise a new 'subtribe to accommodate Hotea which according to him had very

little in common with Deroplax.

No doubt *Hotea* has a typical obovate body and a very long head having accuminate clypeus in contrast to *Deroplax* sp. with comparatively elongated oval posture and less elongated head. *Hotea* has a reduced but a distinct peritreme in the metathoracic scent gland ostioles in contrast to almost an indistinct peritreme in the metathoracic scent gland ostioles of *Deroplax* sp. In the present species of *Hotea* the sepermathecal duct is uniformly thin and remarkably elongated without a median dilation in contrast to a somewhat developed median dilation in *Deroplax* sp.

On the other hand Scudder⁶⁹ has found in D. redtenbacheri and in H. curculionoides a similar form of spermatheca with a very long duct as reported only in Hotea species in the present studies. Finding a very peculiar form of spermatheca in H. subfasciata, Scudder (op. cit.) has suggested the placement of H. curculionoides and H. subfasciata in two different genera.

Probably there is much variation in the group because the group is a primitive one with some members adapting rapidly in accordance with their peculiar habitus. In the entire group, the median dilation is never developed as in pentatomids with sclerotized rod and in some species of *Hotea* (Kumar²⁹) the distal flange is also lacking with a reduced pump region in the spermatheca. Contrary to Kumar²⁹ the present studies revealed three pairs of conjunctival appendages in *Deroplax* sp. similar to those in *Acantholomidea porosa* (Germar) reported by McDonald⁵²

Scutellerinae: This is the largest subfamily and in the present studies also five genera and seven species are included. Pendergrast's⁶¹ material of five species of the family Scutelleridae included four from this subfamily. Kumar²⁹ has also included representatives of eleven genera in the tribe Scutellerini alone in addition to one genus of Elvisurini and one genus of Sphaerocorini. McDonald⁵¹ although studied only one species of Scutellerini, previously he included thirteen species with reference to his studies on male genitalia^{49'50} and nine species with reference to female genitalia⁵¹ (in both *T. diophthalmus* was erroneously included in Scutellerini).

The members of this subfamily show rather stable and somewhat advanced characters. All have relatively a broader head with less developed mandibular plates; all have a well developed metathoracic scent gland ostioles with highly developed peritreme; almost all have well developed parameres with erect stem and a curved and lunate blade with tuft of hairs on the inner basal side of blade; all have thecal processes; in all, the dorsal conjunctival lobes are complicated, and in majority of species the ejaculatory reservoir is greatly developed and in some this supports the vesica.

In females also the eighth paratergites are quite developed (a pentatomid character) and in the spermatheca the pump region with distal and proximal flanges are well developed and usually there is an ovoid or spherical sclerotized and pigmented median dilation which in some (*Chrysocoris*) has ac-

TABLE 3. COMPARATIVE CHARACTERS OF THE METATHORACIC SCENT GLAND OSTIOLE.

				Peritreme	12	1	-	Evaporat	toria
Species	Ostiolar shape	Colour in com- parison to eva- poratoria		Anterior, posterior and outer margins	Form of apex	Transverse length in proprtion to evaporatoria	Colour	Texture	Transverse length i proportion to ter gopleural line
1	2	3	4	5	6	7	8	9	10
E. Integriceps	Elongated, slit- like.	Yellowish with castaneous tips, concolourous.	Elongated.	All sides raised with ant., post., outer and margins sinuate.	Narrowly roun- ded.	Reaching beyond middle.	Yellow.	Faintly rugu- lose.	Reaching one thir distance.
E. maura	Elongated, slit- like.	Castaneous, concolourous.	Elongated.	All sides raised with ant. and post. margins serrate.	Narrowly roun- ded.	Reaching on to the middle.	Black.	Dull velvety.	Reaching closer to
n A.lixoides L and D. zahidae	Deroplax sp. metath	oracic scent gland	d ostioles indisting	ct or absent.					
H. curculionsides	Elongated, slit- like.	Pale yellow, contrasting.	Very small	Posterior margin slightly raised.	Subacute, slightly curved anteriad.	Too small to com- pare.	Ochraceous.	Smooth with black punctures.	Reaching near.
H. nigrorufa,	Very small.	a	b	s e	n	t	Yellowish with reddish tinge.	Rugulose with black punctures.	Reaching near.
5. rubropuncta- tum	Large, wide and ovate, located at apex of vestibular. hump.		it, vestibular hum	np prominently develop	ed.		Yellowish with reddish tinge.	Dull, covered with silvery hairs.	Reaching near.
C. ocellatus	Large, somewhat elongated.	Metalic green, somewhat con- colorous.	Enlarged paddle-like.	Margins faint.	Broadly trun- cated.	Slightly beyond middle.	Light metallic green.	Rugulose, fur- nished with silvery hairs.	Reaching near.
C. stockerus	Rounded	Palish-yellow, contrasting.	Greatly elon- gated, leaf-like.	Posterior and outer margins raised, ant. ter less prominent.	Less broadly rounded with apex curved an- teriad.		Black.	Rugulose and wrinkled.	Reaching on to the middle.
P. latus	Ovate.	Yellowish, only little contrast- ing.		Posterior and outer margins raised, an- terior less promi- nent.		Much beyond middle.	Palish.	Dull and smooth.	Reaching beyond middle.
Poecilocoris p.	Large and ovate.	Yelowlish-brown, concolorous.	Remarkably elongated, thin strip like.	Raised from all sides.	Broadly rounded.	Reaching middle.	Yellowish brown.	Rugulose.	Reaching near.
. fasciata	Rounded.	Palish-yellow, contrasting.	Greatly enlar- ged, leaf-like.	Posterior and outer margins raised, an- terior less prominent.	Broadly rounded.	Much beyond middle.	Bluish black.	Shining, cove- red with silvery hairs.	Reaching one third distance.
. nobilis	Ovate, slit-like	Palish-yellow, contrasting.	Greatly enlar- ged, leaf-like.	Raised from all sides.	Broadly rounded.	Far beyond middle.	Bluish black.	Rugulose.	Reaching closer to.
P.) interruptus Vestwood (but robably another enus)		Metallic bluish black, contrast- ing.		Raised from all sides, post. margin less prominent.	Acuminate, sub- acute, and turned anteriad.	Reaching on to evaporatoria.	Black.	Dull and rugu- lose.	Reaching beyon middle.

I. AHMAD and S. MUSHTAQ

	Pygo	Pygophore		phore Paramere		na		Endosoma			
Species	Ventro-posterior margin and tongue		Stem	Blade	Ventral thecal appendages	Dorsal thecal appendages	Vesica and tip of ejaculatory duct	Dorsal membra- nous conjunctival appendages	Dorsolateral con- junctival appen- dages		
1	2	3	4	5	6	7	8 。 •	9	10		
E. Integriceps	Smooth, medially curved.	Arched and depressed.	Comparatively short.	Very short, T-like.	Absent	Absent	Membranous en- closing ejacula- tory duct.	Paired, medially fused with sclerotized apices.	Paired, sclerotized, elongated, cutely pointed and sword- like.		
E. maura	Sinuate, medially curved.	Arched.	Elongated.	Short, T-like.	Absent	Absent	Membranous, en- closing ejaculatory duct.	Paired, fused at base with semi- sclerotized apices.	Paired, sclerotized, elongated, acutely pointed and sword- like.		
A. tixoides	Slightly sinuate, medially convex.		Short, outer margin sinuate.	Lunate, without- tuft of bristles.	Long, paddle- shaped with truncated apex.		Sclerotized, enclo- sing ejaculatory duct.	Membranous, bil- obed, smoothly tapering apically and attached only at base.	zed, apically lobe-		
Deropalx sp.	Sinuate, medially notched.	Produced into upturned sub- acute spines.	Small, outer margin strongly sinaute.	Very small apex broadly acute with sharp and thick bristles near	Absent	Absent	Membranous, tip of ejaculatory duct coming out.	Largely membra- nous, trilobed, me- dian lobe thickly	Paired but fused throughout largely membranous, with sclerotized tips et		
				hase.				sclerotized.	both sides.		
Males of D.	zahidae, H. curci	ulionoides and	H. nigrorufa wei	e not available.							
S. rubro puncta- tum		Acutely and conspicuously projected, in- ward with knob-like pro- jections.	Short, bent, ou-		Long, tapering distad and api- cally serrate.	Small, fused knob-like in the middle.	Sclerotized, stout with inner margin serrate and tip oot- hed, enclosing eja- culatory duct.	Large sclerotized, paired, elongated with trilobed mem- branous apical por- tions, attached only at base.	dially, very small, entirely membra		
C. ocellatus	Sinuate with me- dian bilobed pro- jection, laterally rounded.		Elongated and erect.	Stout, vertical at base with tuft of bristles.	Knob-like, fused with swollen base and acute apex	Small. fused knob-like in the middle.	Y-shaped, expos-	Membranous pai- red with sclerotized rounded tips and fused only at base.	rounded, lobe-like		
C. stockerus	Sinuate, medially concave.	Lobed	Elongated and erect.	Vertical but curved with a tuft of bristles at base.	Long, slender and horn-like.	Paired, long curved with wide apex.	what stout, in- wardly curved at apex, enclosing tip	Largely sclerotized, membranous only in middle, paired, fused only at base, with horn-like acute apices.	Indistinct.		
P. latus	Medially, V-sha- ped truncated at sides.		Elongated and erect.	Vertical, curved with bilobed apex and thin bristles at base.	Long and uni- formly slender.	Paired, short slender but prominent.	Sclerotized, slen- der, inwardly cur- ved, enclosing eja- culatory duct.	Large sclerotized paired, paddle-like with membranous lobe-like apical portion only at base.	absent.		
Poecilocoris	Medially, V-sha- ped truncated at		Flongated and erect.	Vertical, long bilo- bed at apex, inner	Long and uni- formly slender.	Paired, short slender but prominent.	Sclerotized, slen- der, inwardly cur- ved, venclosing eje-		Indistinct or absent.		

TABLE 4. COMPARATIVE CHARACTERS OF MALE GENITALIA.

× 6

continued

TABLE 4. COMPARATIVE CHARACTERS OF MALE GENITALIA.

1	2	3	4	5	6	7	8	9	10
(P.) interrup tus Westwood (but probably another genus)	Sinuate, medially concave.	Without pro- minent knobs.	Elongated and erect.	Small, vertical, curved with tuft of bristles at base.	Remarkably long and uni- formly slender.	Paired, very short but prominent.	Sclerotized, slender but more stout, enclosing ejacula- tory duct.	dobut more elongated and stout.	Indistinct or absent.
S. fasciata	Emarginate, me- dially concave and laterally hooked.	Smoothly arche with rounded tips.	d Elongated and erect.	Small, lunate a base with tuft o bristles.	t Fused, small- f collar-like in the middle.	Short, slender and curved.	Sclerotized, stout with sinuate lateral margins, enclosing coiled wriglish ejaculatory duct.		
S nobil is	Males not availabl	le.							·

TABLE 5. COMPARATIVE CHARACTERS OF FEMALE GENITALIA.

	7th abdomina	al sternum		Oth and Oth	2		Spermat	h	and the second second
Species	Posterior	Posterolateral	First gonoxae	8th and 9th paratergites					
	margin	tips			Bulb	Pump	Distal flange	Proximal flange	Median dilation
1	2	3	4	5	6	7	8	9	10
								•	
E. integriceps	Medially arched.	Subacute	Moderately concea- led, with straight parallel sided inner margins and rounded but with subacute apices.	extending posteriad	Rounded	Small	Prominent	Prominent	Thickly sclerotized.
F. maura	Medially arched.	Subacute		8th with posterior margin substraight extending posteriad to 9th.	Bulbus	Small	Prominent	Prominent	Thickly sclerotized
	Medially deeply but smoothly arched.	Subrounded .	with straight inner	8th pushed laterad by much larger 9th into strip-like scle- rites.	Bulbus	Small but distinct	Indistinct	Distinct	Indistinct
Female of Der	oplax sp. not ava	ilable for studies	5.						
	Smoothly, medially arched.	Subacute	led, inner margins	8th pushed laterad by much larger 9th into small sclerites.	Small, semi- circular	Well developed	Prominent	Prominent	Dumble-shaped and transparent.
	Smoothly medially arched	Indistinct	Only little concea- led, conical, with straight inner and posterior margins and acute apices.		Oval	Prominent	Small	Small	Absent

TABLE 5. COMPARATIVE CHARACTERS OF FEMALE GENITALIA.

continued

1	2	3	4	5	6	7	8.	9	10
H. nigrorufa	Smoothly medially arched.	Indistinct	dobut posterior margins medially concave.	-do-but into re- latively larger scle- rites.	Elongated	Prominent	Small	Small	Absent
r. rubro- punciatum	Somewhat sinuate laterally but me- dially concave having slight coni- cal projection.		Largely concea- led with straight in- ner and sinuate pos- terior margins with rounded apices.	8th with posterior margins convex, me- dially apart, extend- ing posteriad to eq- ually developed 9th.	Elongated	Small	Distinct	Distinct	Rounded, thick walled.
C. ocellatus	Sinuate, medially deeply emarginate.		Largely exposed, somewhat conical with sinuate poste- rior margins and sub- rounded apices.	8th very large, me- dially apart, extend- ing behind strip-like	Bilobed with apical spheri- cal portion.	Well developed	Distinct	Distinct	Baloon-like, thick-walled.
. stockerus	Slightly laterally sinuate, medially arched with minute median projection			8th triangular, inner tips remarkably nar- rowed, meeting 9th posteriad.	Elongated but apically spheri- cal.	Well developed	Distinct	Distinct	Baloon-like, transparent with inner sclerotized rod.
. latus	Slightly laterally sinuate and me- dially concave.		Partially concealed, inner margins straight posterior margins subtruncate with subrounded apices.		Elongated but apically spher- ical.	Well developed	Distinct	Distinct	Short, rounded and thickwalled,
oecilo- oris sp.	Slightly laterally sinuate, medially arched.	Subrounded	Only little concealed, inner margins con- vex, with broad apices.	-do-but somewhat smaller than 9th.	fusi-form, faintly constricted.	Well developed	Distinct	Dinstinct	Baloon-like, compressed at side, thick-walled.
. fasciatu	Deeply concave in middle.	Rounded	Largely exposed plate-like, tips rounded.	8th with posterior margin elongated and slightly short of 9th, latter small, lobe-like.	Elongated but apically spherical.	Prominent	Distinct	Distnct	Spherical, thick walled.
nobilis	Concave in middle.	Acute	Largely exposed, conical, tips suba- cute.	8th with posterior margin much larger than and extending behind 9th. Latter very small.	Elongated but apically spherical.	Prominent	Distinct	Distinct	Spherical, thick-walled.
y inter- plus West- ood (but robably an- ther genus)	Smoothly arched with faint roun- ded median pro- jection.	Subacute	Only little concea- led, inner margins straight, posterior nearly straight, sub- rounded.	margins convex, extending much be-	Spherical	Well developed	Prominent	Prominent	Baloon-like, thick-walled

EXTERNAL MORPHOLOGY OF Chrysocuris stockerus (LINN.)

dsf

em

em1

em₂

emb

ep

cp1

 ep_2

ev

f1

f2

fe

frt

fur

ge

gp

gu

ip

lbg

lbr

M

map

mdms

mb

mf

mnd

mnl

mnst

mtf

mxl

mxp

s.sp. d.

lst gcx

fam

eye

ds.sp.d

dt.ap

quired the pentatomid-type of sclerotized rod.

McDonald⁵¹ considered the characters of female genitalia of little importance in separating the tribes and subtribes. On the basis of male genitalia, however 49'50 the above author separated Sphaerocorini and Elvisurini from Scutellerini and T. diophthalmus of Pachycorinae according to himself stood quite apart. In Sphaerocorini the above author found strigil on ventral margin of pygophore whereas this character was not found in any member of Scutellerinae.

In Elvisurini the above author found dorsal conjunctival appendages very long and membranous, divided into a dorsal and ventral appendages. The present studies also separated the elvisurine Solenostethium from the rest of the Scutellerini on the basis of characters of the scent gland ostioles and male and female genitalia as noted in the Tables 3-5. Similarly the scutellerine genera are also separated on the basis of these characters. Chrysocoris and P(P) interruptus appear to be more advanced than Scutellera, Poecilocoris and Cantao of the tribe Scutellerini and Solenostethium of Elvisurini as noted above in the characters of the median dilation of spermatheca clearly having pentomid type of sclerotized rod and convex form of venter of abdomen without furrow or sulcation.

Acknowledgement. Dr. Zahida Rishi, Miss M. A. Rizvi and Mr. Mohammad Afzal of the department of Zoology, University of Karachi are acknowledged for technical help and Mr. M. Muneeruddin for typing the manuscript.

The work was supported financially by PL-480 USDA No. A-17-ENT-37 Research Project (FG-Pa-181).

Explanation of Abbreviations

A.4 /A	predetion of theory that to his	P
		mxst
al	anterior laterale	0
an	antenniferoustubercle	oc
anc	antecoxale	occ
anp	anterior notal wing process	ocf
ap	apodeme	ocl
b	bucculae	р
bcs	basicostal suture	par
bl	blade	per
bp	basal plate	pnt
bst	basisternum	ро
cas	circum antennal sulcus	pos
cc	clypeal clefts	pp
cf	claval furrow	pps
cls	claws	pr
cly	clypeus	pra
со	corium	prr
conx	connexivum	ps
cos	circum ocular sulcus	psc
cox	coxa	8th pt
C+Sc	costosubcostal vein	9th pt
Cu	cubitus	pul
cvl	clavus	pxf
cxc	Prothoracic coxal cavitity	px. sp.d.
exc ₁	meso coxal cavity	px.s.sp. d
cxc ₂	metacoxal cavity	
dmca	dorsal membranous conjunctival	ру
	appendages	R

distal flanges distal spermathecal duct dorsal thecal appendage epimeron epimeron of mesothorax. epimeron of metathorax embolium episternum episternum of mesothorax episternum of metathorax. evaporatoria eye frons first fold second fold furcal arm femur furcasternum furca 1st gonocoxae genae gonopore gula inflatory pump labial groove labrum median mandibular plates membrane mandibulo-maxillary suture mesofurca stylet base mandibular lever mandibular stylets metafurcae maxillary lever maxillary plate maxillary stylets ostiole occiput occipital condyles occipital foramen ocelli pygophore paramere peritreme post notum post occiput post occipital sulcus prepectus parapsidal sulcus pleural ridge prealar arms proctiger pleural sulcus prescutum eighth paratergites ninth paratergites pulvillus proximal flanges proximal spermathecal duct swelling of the proximal spermathecal duct. pivot radius

EXTERNAL MORPHOLOGY OF Chrysocoris stockerus (LINN.)

R+M	radio-median vein
sc	sterna costa
scf	semi-circular furrow
scl	supra coxal lobes
sct	scutum
sctl	scutellum
sg 1-4	labial segment 1 to 4
sm. sp.d.	sclerotized median spermathecal
on of the	duct.
sp	spiracle
spb	spermathecal bulb
sp.d.	spermathecal median dilation
sp.p	spermathecal pump
8th. spr.	eighth spiracle
SSS	scuto-scutellar sulcus
st	stem
tap	terminal funnel-shaped apodeme
tar	tarsus
tb	trichobothria
th	theca
tib	tibiae
tibs	tibial spine
tr	triangulin
trc	trochanter
trr	transverse ridge
trs	tarsal spines
ugt	unguitractor plate
ves	vesica
vet	vestibule
vlt	veinlets
vt.ap.	ventral thecal appendage
wng	wing

References

- 1. Q. A. Abbasi and Z. Rishi, Pakistan J. Zool., 5.(2,),189 (1973).
- 2. I. Ahmad and Q. A. Abbasi, J. Sci. Univ. Karachi, 2,(42),60 (1974).
- I. Ahmad and Q. T. Abbasi, Pakistan, J. Zool., 3,(1),37 (1971).
- 4. I. Ahmad and M. U. Shadab, J. Sci. Ind. Res., 18, 133 (1976).
- 5. I. Ahmad and T.R.E. Southwood, Tijdschr. Entomol, 107, 365 (1964).
- S.S. Akbar, Aligarh Muslim Univ. Publ. (Zool. Ser.) Ind. Inst. Type, 5, 1 (1957).
- 7. S. S. Akbar, Aligarh Muslim Univ. Publ. (Zool. Ser.) Ind. Inst. Type., 5(2), 1 (1958).
- 8. L. S. Banerji, Agra Univ. J. Res. Ser., 9, 47 (1960).
- F. Beccari and G. A. Fenili, Rodia Florence, 45, 2, 279 (1961).
- 10. L. Bonnemaisow, Ann. Epiphyt., 2, 127 (1952).
- P. F. Bonhag and J.R. Wick, J. Morphol., 93, 177 (1953).
- 12. F. H. Butt, Corn. Univ. Agr. Expt. Sta., 254, 1 (1943).
- W. E. China and N. C. E. Miller. Bull. Brit. Museum Entomol, 8,(1),1 (1959).
- 14. R. H. Cobben, Centre for Agricultural Publishing and documentation, Wageningen. (1968).
- F. P. Crampton and I.B. Sprague, J. Morphol., 108, 287 (1961).

16. W. L. Distant, Rhynchota., 1 (1902).
17. E. M. Duporte, J. Morphol., 79, 371 (1946).
18. C. Dupuis, Mem. Mus. Hist. nat. Paris (Ser.
A. Zool. 6), 4, 183 (1955).
19. C. Dupuis, Natur. Mus. Hist. Nat. Paris (2),
1 (1963).
20. D. M. Fedotov., Vrednaya cherepachka., 1-3,
Moscow (in Russian). (1947).
21. R. M. Fox and J. W. Fox Reinhold Pub-
lishing Corporation, New York (1964).
22. A.P. Gupta, Tijdschr. Entomol., 106, 169 (1963).
23. M.A. Hamilton, Proc. Zool. Soc. London,
1931, 1067 (1931).
24. R. Khanna, Indian Tr. Entomol., 25, 63 (1963).
25. H. H. Knight, Bull. Ill. Nat. Hist. Serv., 22.(1), 1 (1941).
26. R. Kumar, Entomol, Tidskr., 83, 44(1962a).
27. R. Kumar, Zoologica Poloniae., 12 (1), 3 (1962b).
28. R. Kumar, Proc. Royal Soc. Queenland.,
75. 8, 51 (1954).
29. R. Kumar, J. entomol. Soc. Queensland, 4, 41
(1965).
30. O. Larsen, Lund. Univ. Arsskr. N. F. Ave.,
2, 41 (11), 1 (1945a).
31. O. Larsen, Lund. Univ. Arsskr. N. F. Ave.,
2 , 41 (3), 1 (1945b).
32. J. D. Lattin, Ph. D. Thesis, Univ. of California,
Berkeley (1964).
33. D. R. Lauck, Ann. Entomol, Soc. Am., 52, 93
(1959).
34. D. Leston, Cult. Co. Diam. Angola., 6, 9 (1952).
35. D. Leston, Ent. Gaz. London., 4, 13 (1953a).
36. D. Leston, J. Soc. Brit. Entomol., 4, 120 (1953b).
37. D. Leston, Rev. Brazil. Biol., 13, 121 (1953c).
38. D. Leston, Proc. R. ent. Soc. London (A)., 29,
9 (1954).

- 39. D. Leston, Proc. Zool. Soc. London, **128**, 369 (1957).
- 40. D. Leston, Proc. 10th. Int. Cong. Ent. Montreal 1959, 325 (1958a).
- 41. D. Leston, Int. Congr. Ent. 10. Montreal 1959., 2, 911 (1958b).
- D. Leston, Proc. R. ent. Soc. London, 37 (10-12), 135 (1962).
- 43. N. Lodos, Psyche. Cambridge. Mass., 70, 144 (1963).
- 44. E. I. MacGill, Proc. Zool. Soc. London, 117, 1067 (1947).
- 45. N. S. R. Malouf, Bull. Soc. Royal Entomol. Egypte. n.s., 16, 161 (1933).
- 46. E. P. Marks, J. Kans, Entomol. Soc., 24, 134 (1951).
- 47. R. Matsuda, Mem. Am. Entomol. Inst., 4, 1 (1965).
- 48. R. Matsuda, Mem. Entomol. Soc. Canada. No. 76, 1 (1970).
- 49. F. J. D. McDonald, Pap. Dept. Entomol. Univ. Queensland, 1,(12),173 (1961).
- F. J. D. McDonald, J. ent. Soc. Qd. Brisbane..
 2, 24 (1963a).
- 51. F. J. D. McDonald, Univ. Qd. pap. Brisbane (Entomol), 1(15), 229(1963b).
- 52. F.J.D. McDonald, Quaest Entomol., 2, 7 (1966).
- 53. S. Miyamoto, Sieboldia, 2, 197 (1961).

- 54. F. Muir and J. C. Kershaw, Psyche. Cambridge Mass., 18, 1 (1911).
- 55. J. G. Myers, Proc. Zool. Soc. London, 1928, 365 (1928).
- 56. W. S. Newcomer, J. Morphol., 82, 365(1948).
- 57. P. Nuorteva, Ann. ent. Fenn., 22, 45 (1956).
- 58. M. C. Parsons, Bull. Mus. Comp. Zool. Merr. Coll., 122, 1 (1959).
- 59. M. C. Parsons, Bull. Mus. Comp. Zool. Merr. Hom. Coll., 122,(7), 299 (1960).
- 60. M.C. Parsons, Trans. R. Entomol Soc. London, 114, 97 (1962).
- 61. J. G. Pendergrast, Trans. Royal Entomol. Soc. London, 109(1), 1 (1957).
- 62. F. Piotrowski, Soc. Amis. Sci. Lettr. Poznarc (E), 12, 237 (1950).
- 63. H. S. Pruthi, Trans. Royal Entomol. Soc. London, 78, 127 (1925).
- 64. M. A. H. Qadri, Proc. Zool. Soc. Bengal, 2 (1),43 (1949).
- 65. M.A.H. Qadri and S.A. Aziz, Aligarh Muslim Univ. Publ. (Zool. Ser.) Ind. Ins. Typ., 2, 1 (1950).
- 66. S. A. Qureshi, I. Ahmad and S. N. H. Naqvi, Folia Biologica, 19,(3),385 (1971).
- 67. B.L. Rawat, Trans. Royal Entomol. Soc. London, 88, 119 (1939).
- 68. C. W. Schaefer, J. N. Y. Entomol. Soc., 76,(2), 87 (1968).
- 69. G. G. E. Scudder, Trans. Royal Entomol. Soc. London, 111, 405 (1959).

- 70. R. E. Snodgrass, McGraw-Hill Book Company Inc. New York and London (1935).
- 71. R. E. Snodgrass, Smithsonian. Misc. Coll., 135, 6, 1 (1957).
- 72. R. E. Snodgrass, Smithsonian Misc. Coll., 140, 1, 1 (1960).
- 73. T. R. E. Southwood, Trans. Royal Entomol. Soc. London, 104, 415 (1953).
- 74. T. R. E. Southwood, Tijdschr. Entomol. 9. 77 (1955)
- 75. T. R. E. Southwood, Trans. Royal Entomol., Soc. London, 108, 163 (1956).
- 76. T. R. E. Southwood and D. Leston, Frederick Werne and Co. Ltd. London and New York (1959).
- 77. C. S. Spooner, Illinois Biol. Monogr., 1 (1938). 16.
- 78. P. Stys, Verh. XI. Int. Mongr. Entomol Wien. 225 1960. Vienna., 1, 37 (1962).
- 79. P. Stys, Acta. Zool. Acad. Sci. Hungar., 10, 229 (1964).
- 80. L. M. Taylor, Ann. Entomol. Soc. Am., .11, (1918).
- 81. D.G. Tower, Ann. Entomol. Soc. Am., 6, 427 (1913). 82. S. L. Tuxen, ed., Ejnar Munksgaard, Copen-
- hagen, 284 pp.
- 83. I. Vidal, Mem. Soc. Nat. Mar., 48, 1 (1949).
- 84. E. Wagner, Beitr. Z. Entomol. I. III., 14, 383 (1951).
- 85. E. Wagner, Acta. Entomol Mus. Nat. Pragae., 35, 73 (1963).