

STUDIES ON STORED GRAIN FUNGI

Part III.—Fungi from Cereals

M.A. AHMED and S. SHAHID HUSAIN

PCSIR Laboratories, Karachi 39

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Thirty-eight species belonging to 15 genera were isolated from *Zea mays* Linn. (Pakistan), *Zea mays* Linn. (American), *Hordeum vulgare* Linn., *Sorghum vulgare* Pers and *Pennisetum typhoideum* Rich stored in various godowns in Karachi. About 83% of the organisms belonged to Fungi Imperfecti while Ascomycetes (15%) and Phycomycetes (2%) accounted for the remaining fungi. Species of *Aspergillus* were most predominant among all the fungi recorded and among these species *A. flavus* was the most prevalent organism. In *Zea mays* (American) damage was most severe because of the high moisture contents and increased temperature which resulted in the formation of "hot spots" and extensive damage. Surface sterilization of grains with 1:1000 HgCl₂ reduced the number of fungi considerably.

The authors have already reported^{1,2} the fungi isolated from wheat and rice and from oilseeds alongwith *Plantago ovata* stored in various godowns in Karachi. Considerable amount of work has been done on fungi inhabiting and infesting cereals alongwith effect of environmental factors, moisture content, temperature in storage etc. on the severity of deterioration of grains. Sinha and Wallace³ studied fungi causing deterioration of wheat during storage. They observed an ecological succession of species of microorganisms, which often overlapped, in the following order: *Penicillium cyclopium*, *P. funiculosum*, *Aspergillus flavus* Link, *A. versicolor*, *Absidia* sp and *Streptomyces* sp. Lutey and Christensen⁴ observed the influence of moisture content, temperature and length of storage upon survival of fungi in barley kernels. Machacek *et al.*⁵ determined the effect of high water content in stored wheat, oat, and barley seeds on susceptibility to invasion by moulds. Qasem and Christensen⁶ carried out investigations on the influence of various factors on the deterioration of stored corn by fungi. Barron and Lichwardt⁷ worked on estimation of fungi associated with deterioration of stored maize. Junejo and Malik⁸ carried out investigations on the microflora associated with sorghum and have reported a number of fungi causing deterioration of the grains.

The present investigation is an effort to determine the fungi infesting and causing deterioration of *Zea mays* Linn., *Hordeum vulgare* Linn., *Sorghum vulgare* Pers. and *Pennisetum typhoideum* Rich in storage.

Materials and Method

The stored grains were collected by random sampling method from the godowns in Karachi at different times of the year during 1969. The grains were stored in jute bags mostly placed on wooden platforms. The grain which were sampled for presence of fungi were *Zea mays* Linn. (Pakis-

tani), *Zea mays* Linn. (American), *Hordeum vulgare* Pers. and *Pennisetum typhoideum* Rich.

The temperature, outside and inside the bags of grain, was also determined. Air oven method⁹ was applied for determining the moisture contents of the grains. For details of the methods reference may be made to our earlier communications.^{1,2}

Results

A total number of 38 species belonging to 15 genera were isolated from the four cereals under investigation. Some organisms were common among all the cereals while some of them were isolated from only one or two cereals. Species of *Aspergillus* formed the nucleus of fungi infesting the cereals in godowns (Fig. 1).

There were two kinds of maize stored in godowns, one grown in Pakistan, the other imported from U.S.A. The one imported from U.S.A. was stored in godowns and some of them had hot spots (a condition where the temperature inside the heap of grains becomes high due to activities of microorganisms and high moisture contents which result in much more deterioration of the grain).

Table 1 lists the fungi isolated from the four cereals under investigation.

Investigations were also carried out to observe the effect of surface sterilization on the grains with respect to emergence of different organisms from them. The grains were surface sterilized with 1:1000 HgCl₂ for different lengths of time. After 10-sec exposure the number of organisms considerably reduced and only three species of fungi from each cereal, with the exception of *Sorghum vulgare* where only two organisms appeared, were isolated. At 20 sec, with the exception of *Hordeum vulgare* where three organisms emerged, the rest of the cereals yielded only 2 species each. When the grains were sterilized for 40 sec, no organism

appeared from *Sorghum vulgare*, two organisms each were isolated from *Hordeum vulgare*, *Zea mays* (American) and *Zea mays* (American burned) while only one fungus each emerged from *Pennisetum typhoideum* and *Zea mays* (Pakistani). At 60 sec no organism appeared from *Pennisetum typhoideum* while one species each was isolated from the rest of the cereals. *Aspergillus flavus* was isolated from *Hordeum vulgare* and *Zea mays* (Pakistani) while *A. niger* appeared in the American maize and no organism was isolated from other cereals when the seeds were surface sterilized for 80 sec. At 100 sec, *A. niger* and *A. flavus* were isolated from the American variety while the rest of the grains failed

to yield any fungi. Table 2 shows the fungi isolated from various cereals after exposure to 1:1000 HgCl₂ for different lengths of time. Figure 2 shows the relative prevalence of fungi after surface disinfection.

Discussion

Fungi Imperfecti was the dominant group with a prevalence of 83%. Phycomycetes and Ascomycetes have a prevalence of 15 and 2% respectively.

The genus *Aspergillus* was the most prevalent organism with 11 species and among the species *A. flavus* topped the list with a prevalence of 22%. *A. tamarii*, however, was the least prevalent or-

TABLE I.—FUNGI ISOLATED FROM CEREALS BEFORE SURFACE DISINFECTION.

<i>Hordeum vulgare</i> Linn.		<i>Sorghum vulgare</i> Pers.	
1. <i>Aspergillus niger</i> van Tiegh		1. <i>Aspergillus niger</i> van Tiegh	
2. <i>A. flavus</i> Link.		2. <i>A. flavus</i> Link.	
3. <i>A. sydowii</i> (Bain and Sart) Thom & Church		3. <i>A. fumigatus</i> Fres.	
4. <i>A. fumigatus</i> Fres.		4. <i>A. nidulans</i> (Eidam) Wint.	
5. <i>A. candidus</i> Linn.		5. <i>Fusarium semitectum</i> Berk & Rav	
6. <i>A. violaceus</i> Raper & Fennel		6. <i>F. chlamydosporum</i> Wr. & Rg	
7. <i>A. nidulans</i> (Eidam) Wint.		7. <i>Grechslera hawaiiensis</i> (Bugn.) Subram & Jain	
8. <i>A. terreus</i> Thom.		8. <i>Trichoderma harzianum</i> Rifai agg.	
9. <i>Alternaria alternata</i> (Fr.) Keissler.		9. <i>Absidia ramosa</i> (Lindt) Lender.	
10. <i>A. triticina</i> Prasada and Prabhu.		10. <i>Alternaria alternata</i> (Fr.) Keissler.	
11. <i>Curvularia lunata</i> (Wakker) Boedijn.		11. <i>Pleospora infectoria</i> Fuckel	
12. <i>Drechslera hawaiiensis</i> (Bugn.) Subram & Jain		12. <i>Cladosporium sphaerospermum</i> Penz.	
13. <i>Cochliobolus spicifer</i> Nelson.			
14. <i>Cladosporium sphaerospermum</i> Penz.			
15. <i>Rhizopus arrhizus</i> Fischer			
16. <i>Penicillium decumbens</i> Thom			
17. <i>P. chrysogenum</i> Thom.			
18. <i>Fusarium equiseti</i> (Corda) Sacc.			
19. <i>Monilia</i> sp.			
<i>Pennisetum typhoideum</i> Rich		<i>Zea mays</i> Linn. (Pakistani)	
1. <i>Aspergillus niger</i> van Tiegh.		1. <i>Aspergillus niger</i> van Tiegh.	
2. <i>A. flavus</i> Link.		2. <i>A. flavus</i> Link.	
3. <i>A. oryzae</i> (Ahlb.) Cohn.		3. <i>A. ochraceus</i> Wilhelm	
4. <i>A. violaceus</i> Raper and Fennel		4. <i>A. nidulans</i> (Eidam) Wint.	
5. <i>A. nidulans</i> (Eidam) Wint.		5. <i>A. tamarii</i> Kita.	
6. <i>A. ochraceus</i> Wilhelm.		6. <i>A. fumigatus</i> Fres.	
7. <i>Penicillium notatum</i> Westling.		7. <i>Penicillium notatum</i> Westling.	
8. <i>P. brevicompactum</i> Dierckx.		8. <i>Rhizopus arrhizus</i> Fischer	
9. <i>Rhizopus arrhizus</i> Fischer		9. <i>R. oryzae</i> (Went & Geerl)	
10. <i>Alternaria alternata</i> (Fr.) Keissler.		10. <i>Trichoderma polysporum</i> (Link ex Pers) Rifai	
11. <i>Curvularia lunata</i> (Wakker) Boedijn.		11. <i>Fusarium moniliforme</i> Sheld.	
12. <i>C. penniseti</i> (Mitra) Boedijn.		12. <i>Neurospora sitophila</i> Shear & Dodge	
13. <i>Drechslera hawaiiensis</i> (Bugn.) Subram & Jain.		13. <i>Alternaria alternata</i> (Fr.) Keissler.	
		14. <i>Monilia</i> sp.	
<i>Zea mays</i> Linn. (American)		<i>Zea mays</i> Linn. (with hot spots)	
1. <i>Aspergillus niger</i> van. Tiegh.		1. <i>Aspergillus niger</i> van Tiegh.	
2. <i>A. flavus</i> Link.		2. <i>A. flavus</i> Link	
3. <i>A. violaceus</i> Raper & Fennel		3. <i>A. amstelodami</i> (Mangin). Thom & Church.	
4. <i>Rhizopus arrhizus</i> Fischer.		4. <i>A. violaceus</i> Raper and Fennel.	
5. <i>Penicillium decumbens</i> Thom.		5. <i>A. nidulans</i> (Eidam) Wint.	
		6. <i>A. terreus</i> Thom.	
		7. <i>Penicillium islandicum</i> Sopp.	
		8. <i>P. notatum</i> Westling	
		9. <i>P. decumbens</i> Thom.	
		10. <i>P. citrinum</i> Thom.	
		11. <i>Curvularia lunata</i> (Wakker). Boedijn.	
		12. <i>Cladosporium sphaerospermum</i> Penz.	
		13. <i>Sepedonium</i> sp.	

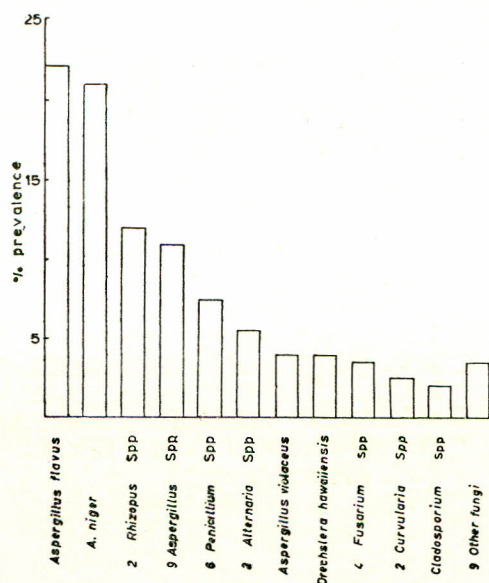


Fig. 1.—Percent prevalence of fungi before surface disinfection.

ganism among the *Aspergillus* spp. with a frequency of 0.3%. Species of *Penicillium* were also fairly dominant as the genus was represented by six species. *P. decumbens* and *P. citrinum* with 2% and 0.30% respectively were the most and the least prevalent fungi. The species of *Penicillium* and *Aspergillus* accounted for almost half the isolated organisms. The rest of the organisms were mostly field fungi and were represented by a few species. As has been observed, temperature plays an extremely important role in the deterioration of stored food grains. Maximum temperature of 37°C was recorded inside the heap of grains of the American variety of maize which had hot spots, while the outside temperature was 32°C. The grains were very badly damaged. The moisture

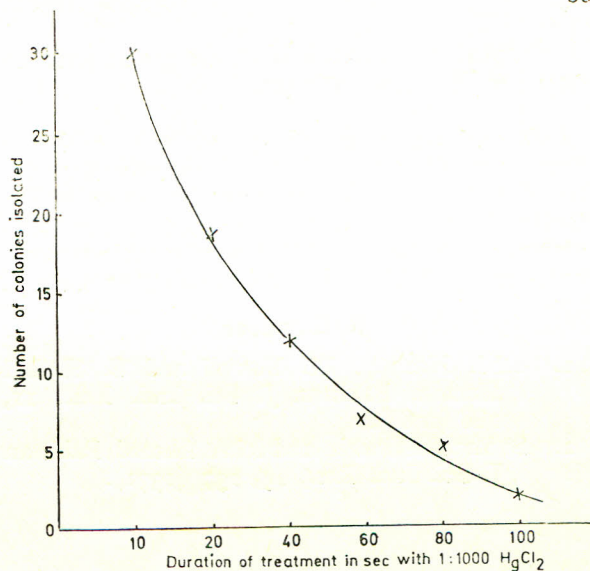


Fig. 2.—The number of colonies isolated after surface disinfection with 1:1000 HgCl₂.

content of the grain in this variety was highest (27%). The temperature of all the other grains was about 34.5°C.

Moisture content, of course, varied greatly: *Pennisetum typhoideum*, 10%; *Hordeum vulgare*, 23%; *Sorghum vulgare* 11%; *Zea mays* (American), 16%; *Zea mays* (with hot spots), 27%; *Zea mays* (Pakistani), 18%.

Although less moisture content is generally associated with decrease in microorganism activity, however in case of *Pennisetum typhoideum* considerable number of fungi were isolated and the damage to the grains was also fairly widespread.

As expected, surface sterilization of grain with 1:1000 HgCl₂ reduced the number of fungi considerably and as may be noted from Table 2 very

TABLE 2.—FUNGI ISOLATED AFTER SURFACE DISINFECTION WITH 1:1000 HgCl₂.

	<i>Pennisetum typhoideum</i>	<i>Hordeum vulgare</i>	<i>Sorghum vulgare</i>	<i>Zea mays</i> (American)	<i>Zea mays</i> (with hot spots)	<i>Zea mays</i> (Pakistani)
10 sec	<i>Aspergillus flavus</i>	<i>Aspergillus flavus</i>	<i>Aspergillus flavus</i>	<i>Aspergillus flavus</i>	<i>Aspergillus flavus</i>	<i>Aspergillus flavus</i>
	<i>A. niger</i>	<i>A. niger</i>	<i>A. niger</i>	<i>Rhizopus arrhizus</i>	<i>A. amstelodami</i>	<i>Rhizopus arrhizus</i>
	<i>Penicillium notatum</i>	<i>Alternaria alternata</i>			<i>Penicillium islandicum</i>	
20 sec	<i>Aspergillus flavus</i>	<i>Aspergillus flavus</i>	<i>Aspergillus niger</i>	<i>Aspergillus niger</i>	<i>Aspergillus niger</i>	<i>Aspergillus flavus</i>
	<i>A. niger</i>	<i>A. niger</i> <i>A. candidus</i>	<i>Alternaria alternata</i>	<i>A. flavus</i>	<i>A. amstelodami</i>	<i>A. niger</i>
40 sec	<i>Alternaria alternata</i>	<i>A. fumigatus</i> <i>Rhizopus arrhizus</i>	—	<i>A. niger</i> <i>Rhizopus arrhizus</i>	<i>A. niger</i>	<i>A. nidulans</i>
60 sec	—	<i>Aspergillus flavus</i>	<i>Aspergillus niger</i>	<i>Aspergillus niger</i>	<i>A. flavus</i>	<i>A. flavus</i>
80 sec	—	<i>Aspergillus flavus</i>	—	<i>A. niger</i>	<i>A. niger</i>	<i>A. flavus</i>
100 sec	—	—	—	<i>A. niger</i>	<i>A. flavus</i>	—

few organisms appeared when the grains were exposed to HgCl_2 for 100 sec.

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